

Morphological classification of parenchymal disorders of the canine and feline liver

3. Hepatic abscesses and granulomas, hepatic metabolic storage disorders and miscellaneous conditions

Tom Van Winkle, John M. Cullen, Ted S.G.A.M. van den Ingh, Jenny A. Charles, Valeer J. Desmet

Update 2021: John M Cullen, Ted SGAM van den Ingh, Guy CM Grinwis, Hille Fieten

ABSTRACT

Hepatic abscesses and granulomas usually occur by hematogenic spread from the portal vein or in neonates from the umbilical vein and a wide variety of causative organism is given. The various hepatic metabolic storage diseases in dogs and cats mentioned in the literature are summarized including the enzymatic defect, the hepatic cells involved and their morphological aspect. Finally, miscellaneous conditions are presented which include cytoplasmic and nuclear alterations in hepatocytes, Kupffer cells and hepatic stellate cells as well as lipogranulomas and pigment granulomas and extramedullary hemopoiesis.

HEPATIC ABSCESSSES AND GRANULOMAS

Hepatic abscesses usually are the result of bacterial infections evoking intense accumulation and subsequent lysis of neutrophilic granulocytes at the infection site (Fig. 1). They can reach the liver via different routes including the portal vein or umbilical vein, ascending infection of the biliary system, and by direct contact and penetration of the liver capsule. Hepatic abscesses in dogs and cats are particularly seen in newborn animals due to umbilical infection (e.g. several Gram-positive and Gram-negative bacteria). In adult animals hepatic abscesses are often the result of infections with *Yersinia spp* (Fig. 2), *Nocardia asteroides* and *Actinomyces spp*.^(1, 2) Hepatic abscesses may occur in association with central necrosis of hepatocellular neoplasms.

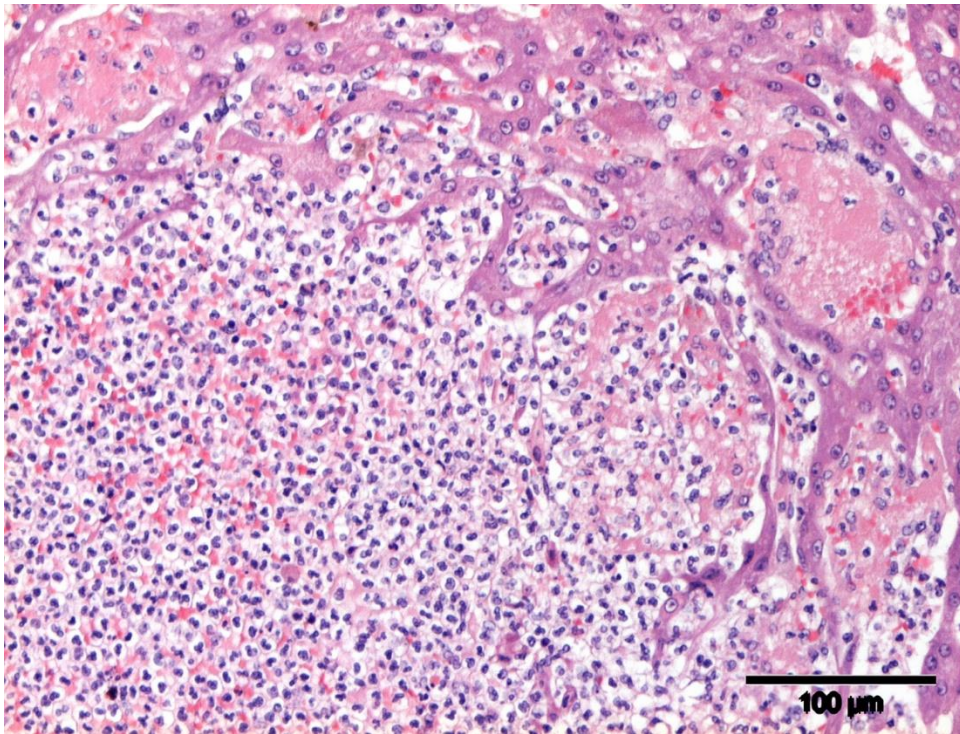


Fig. 1. Dog. Hepatic abscess with fibrinoid deposits at the periphery. HE.

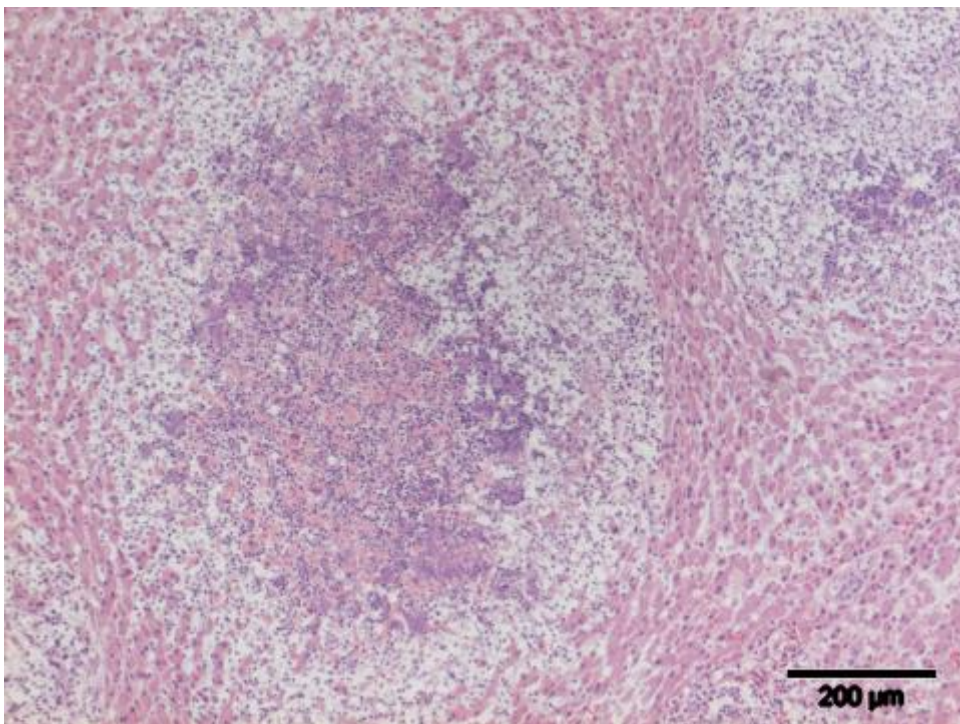


Fig. 2. Cat. Hepatic abscess due to Yersinia spp with centrally located bacteria. HE.

Hepatic granulomas may occur in a wide variety of diseases, some of which are primary in the liver, but most are part of a generalized disease process. They consist of (multi)focal aggregations of activated macrophages with an epithelioid appearance, mostly infiltrated by

lymphocytes and plasma cells and possibly fibroblasts and may be surrounded by collagen fibers. Epithelioid cells may fuse to form multinucleated giant cells; the nuclei may either be arranged peripherally (Langhans-type giant cell) or haphazardly (foreign-body-type giant cell).⁽³⁾ The classic example of granulomatous disease is tuberculosis. The granulomas, referred to as tubercles, are characterized by the presence of epithelioid cells and Langhans-type giant cells with central caseous necrosis and a peripheral zone of lymphocytes (Fig. 3); in contrast to other domestic animal species giant cells of Langhans type are only rarely observed in both dogs and cats⁽⁴⁾.

Infectious causes for hepatic granulomas in dogs and cats include mycobacterial infections (*M. tuberculosis* (Fig. 3), *M. avium intracellulare* (Fig. 5)), systemic mycoses (*Blastomyces dermatitidis*, *Cryptococcus neoformans*, *Histoplasma capsulatum*, *Coccidioides immitis*) and opportunistic fungal infections (Fig. 6), migrating nematode larvae (visceral larva migrans) and schistosomiasis. Alveolar echinococcosis due to *Echinococcus multilocularis* is sometimes seen in dogs in endemic areas and usually presents as one large or multiple neoplastic-like processes in the liver. Histologically the lesion is characterized by the presence of, usually dead, echinococcal cysts embedded and surrounded by chronic granulomatous inflammation. (Fig. 7)

A diffuse granulomatous inflammation of the liver is observed in *Leishmania spp* infection (Fig. 8) in dogs⁽⁵⁾ and *Cytauxzoon felis* infection (Fig. 9) in cats⁽⁶⁾; in both cases large numbers of amastigotes respectively schizonts may be observed in Kupffer cells and macrophages. Granulomatous and non-suppurative hepatitis has been reported in the dog in association with *Bartonella spp.*⁽⁷⁾

Granulomas may also be incited by relatively inert foreign material (e.g. crystalline material, sutures, plant material). In neonates, subcapsular hepatocellular necrosis (probably associated with asphyxia) with subsequent mineralization and granulomatous inflammation may be seen at the periphery of the liver lobes (Fig. 10).

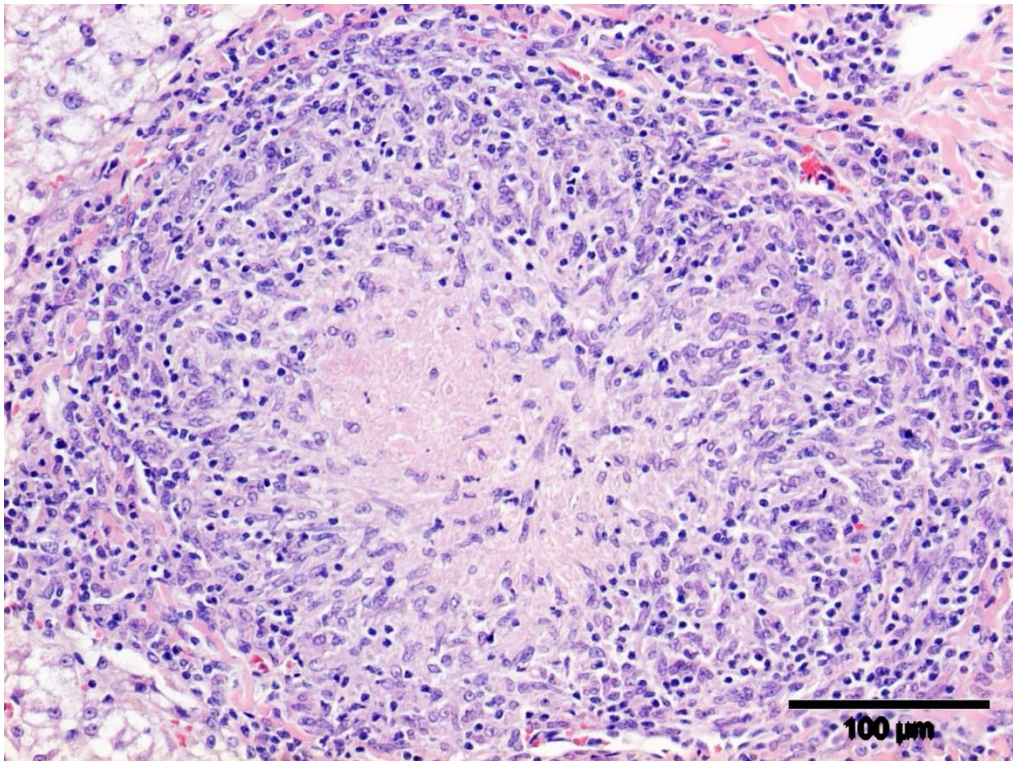


Fig. 3. Dog. Tubercle with epithelioid cells, central caseous necrosis and peripheral lymphocytic infiltrate. Note absence of Langhans type multinucleated giant cells. Mycobacterium tuberculosis. HE.

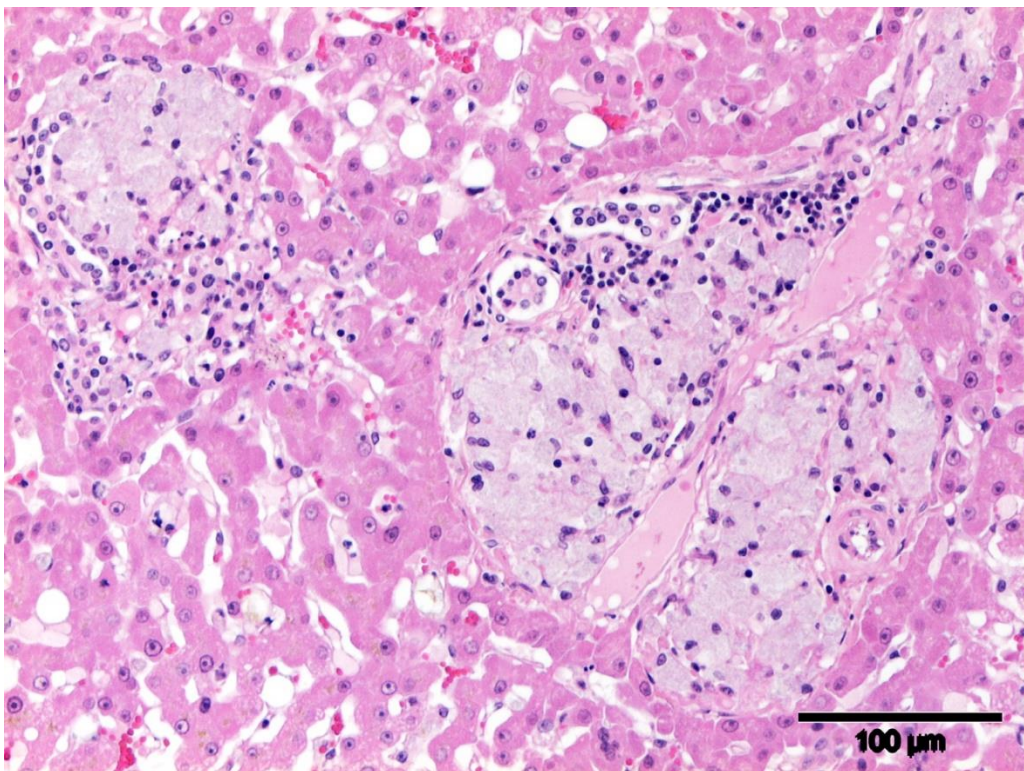


Fig. 4. Cat. Epithelioid granulomas in the portal area and parenchyma. Mycobacterium avium. HE.

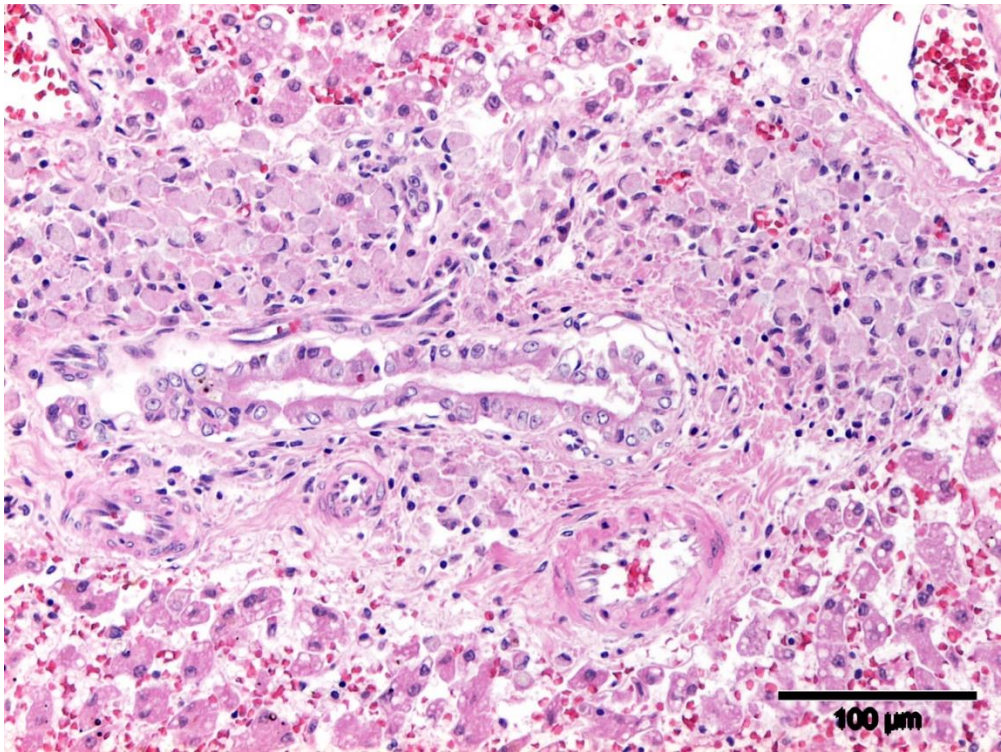


Fig. 5. Dog. Epithelioid granulomatous inflammation in the portal area. Mycobacterium avium. HE.

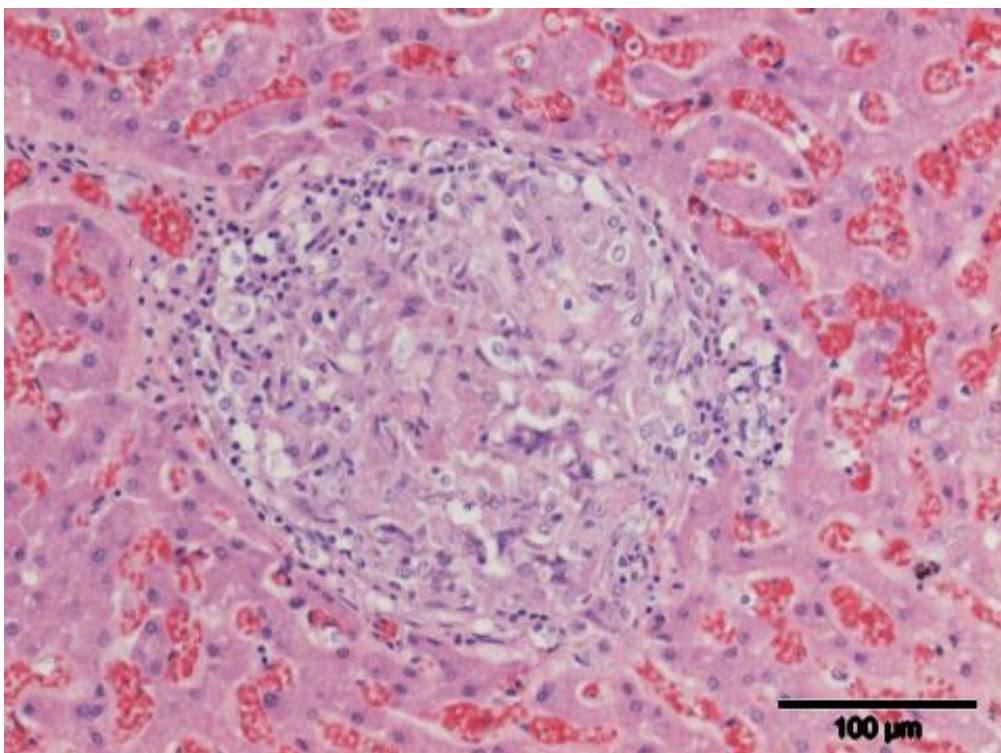


Fig. 6. Dog. Mycotic granuloma with pale irregularly formed fungal organisms. HE.

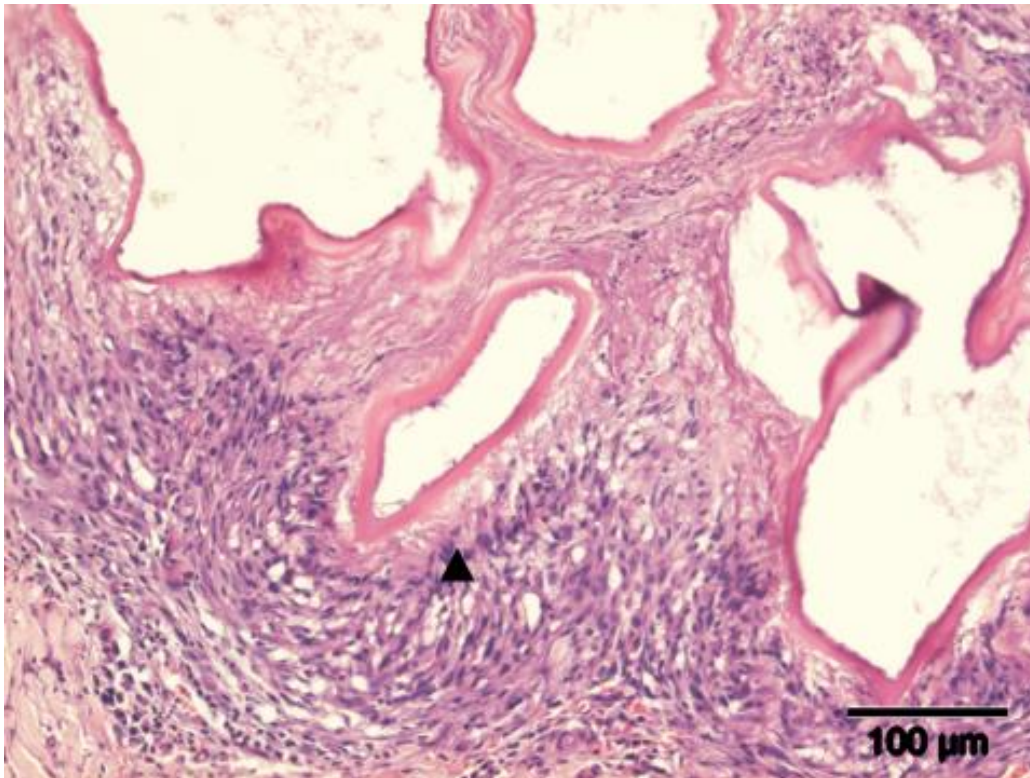


Fig. 7. Dog. Alveolar echinococcosis. Dead echinococcal cysts surrounded by a ring of foreign-body multinucleated giant cells (arrowhead) and chronic inflammation. HE.

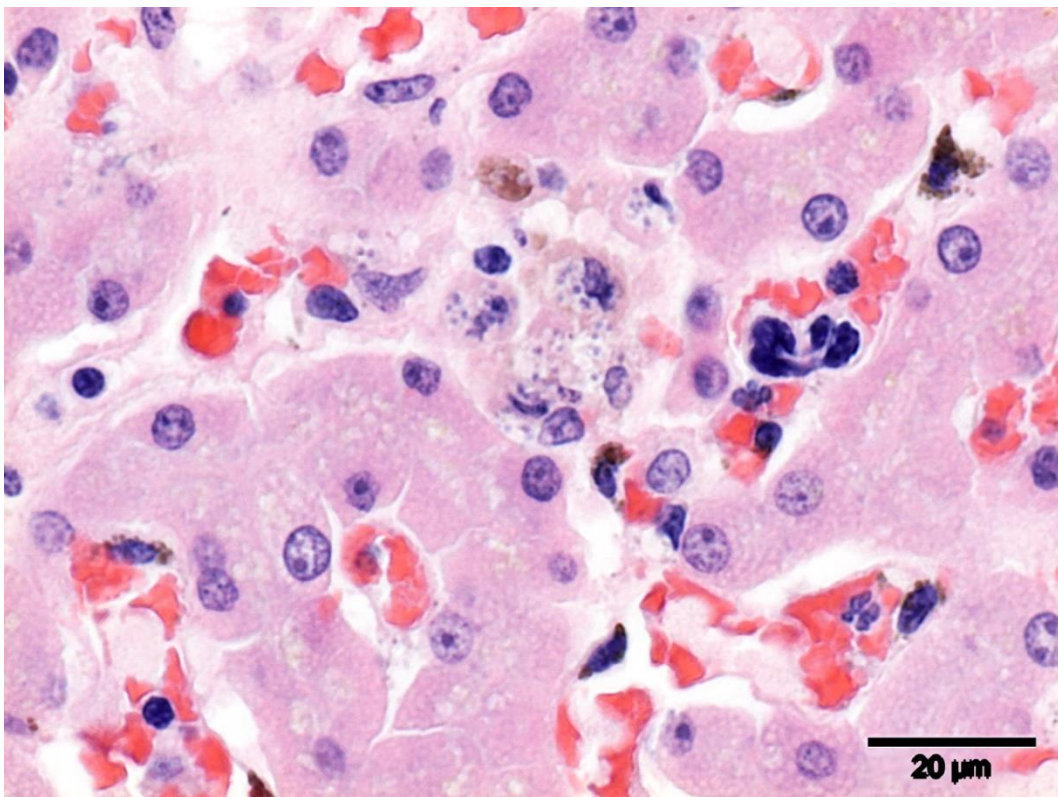


Fig. 8. Dog. Leishmaniasis. Amastigotes in the cytoplasm of Kupffer cells / macrophages. HE.

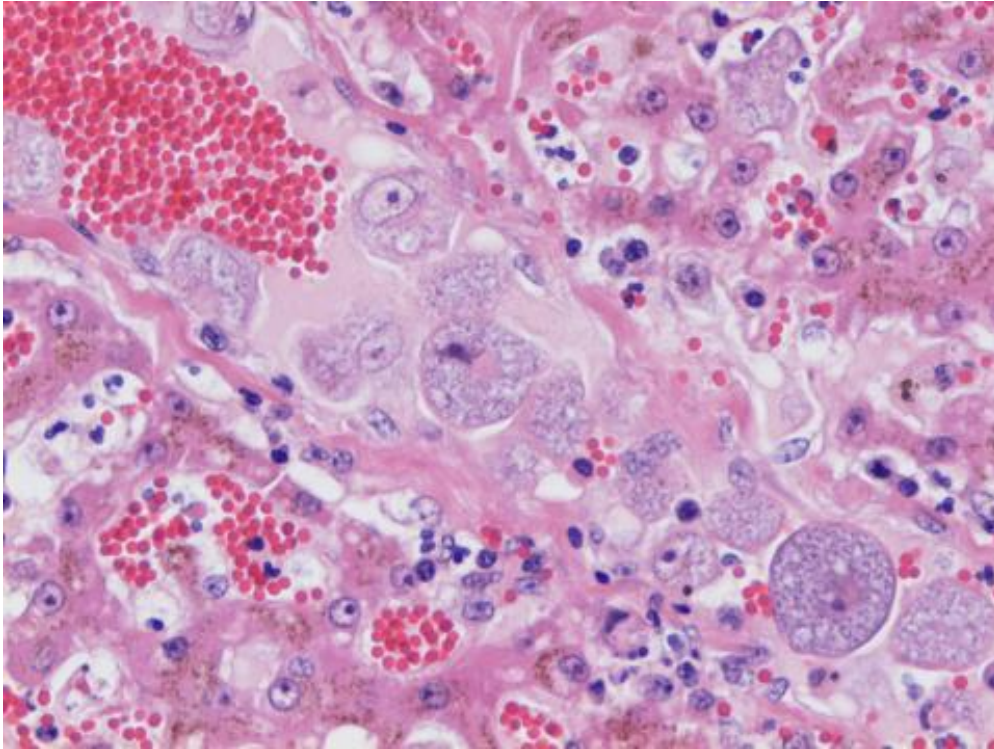


Fig. 9. Cat. Cytauxoon felis infection. Large numbers of schizonts in Kupffer cells and macrophages. HE.

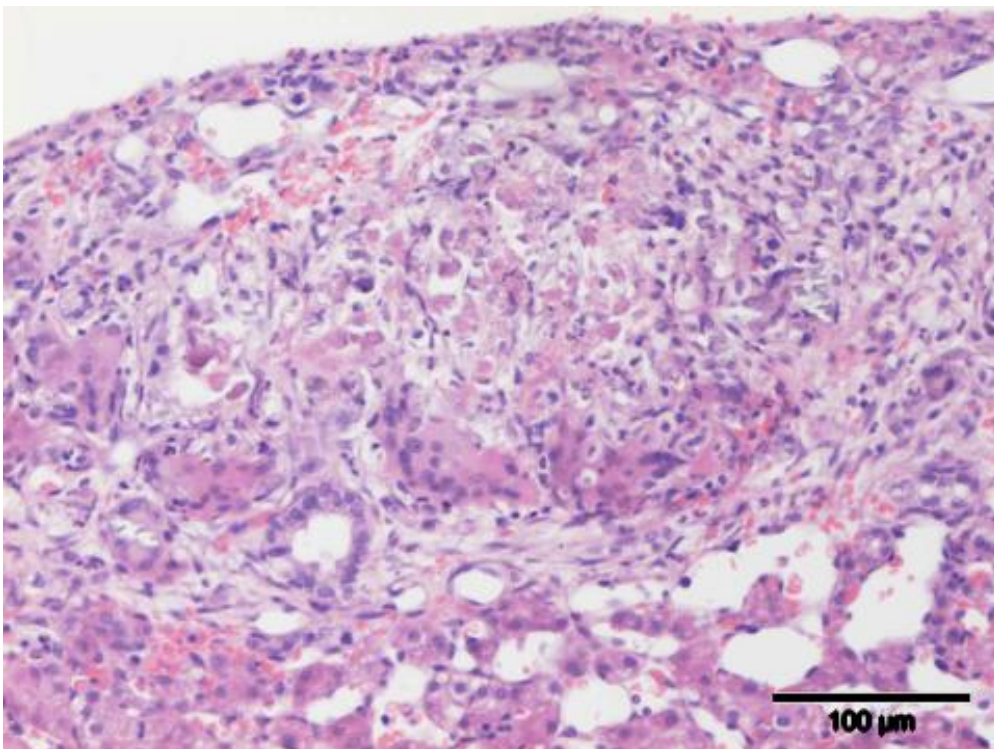


Fig. 10. Dog, neonate. Subcapsular necrosis with dystrophic calcification and subsequent granulomatous inflammation. HE.

HEPATIC METABOLIC STORAGE DISORDERS

General

Hepatic metabolic storage disorders, usually associated with inherited but sometimes acquired metabolic enzyme deficiencies, can have a variety of morphologic appearances (Table 1). The most common finding is the presence of clear vacuoles, vacuoles with granular or hyaline material, or pigmented granules in hepatocytes and, or Kupffer cells and macrophages (Fig. 11, 12). Rarely, as in glycogenosis type I and III, the lesion is characterized by markedly swollen, clear hepatocytes with well outlined cell membranes giving the hepatocytes the appearance of plant cells.

Table 1 Hepatic metabolic storage disorders						
Disease name	Stored material	Defective enzyme	Morphology of stored material	Hepatocytes	Kupffer cells / macrophages	Species / breed
Ceroid-lipofuscinosis	ceroid / lipofuscin	Unknown	granules (yellow-brown)		X	Canine ¹⁵⁻¹⁷ Feline ^{17,18}
Cholesterol ester storage disease	cholesterol esters and cholesterol in Kupffer cells and macrophages; neutral lipids and cholesterol in hepatocytes	acid lipase	clear vacuoles and cholesterol crystals	X	X	Canine ¹⁹ Fox terrier
Fucosidosis	fucose containing glycolipids, glycoproteins, poly- / oligo-saccharides	alpha-L-fucosidase	clear vacuoles		X	Canine ^{20,21} English springer spaniel

Galactosialidosis	glycolipids and oligosaccharides	beta galactosidase, alpha-neuraminidase	clear vacuoles	X	X	Canine ²² Schipperke dog
GM 1 gangliosidosis	gangliosides	beta-galactosidase	clear vacuoles	X	X	Canine ²²⁻²⁸ Portugese waterdog, Shiba dog, Alaskan husky, English springer spaniel, Mixed breed dog Feline ^{29,30} Siamese, Korat
GM2 gangliosidosis	gangliosides	beta-hexosaminidase A / B	clear vacuoles	X	X	Canine ³¹⁻³³ Japanese spaniel, German shorthaired pointer, Golden retriever, Feline ³⁴ DSH
Glycogenosis type IA	glycogen	glucose-6-phosphatase	swollen clear cells	X		Canine ³⁵ Maltese dog
Glycogenosis Type III	glycogen	amylo-1,6-glucosidase	swollen clear cells	X		Canine ^{36,37,63,65} German sheperd dog, Curly-coated retriever
Glycogenosis Type IV	abnormally branched glycogen (alpha-1,4-D-glucan)	branching enzyme	pale blue granules	X	X	Feline ³⁸ Norwegian forest cat

WSAVA Standards for Clinical and Histological Diagnosis of Canine and Feline Liver Diseases
Society of Comparative Hepathology

Glycosyl ceramide lipidosis (Gaucher's disease)	glycosyl ceramines	acid beta-glucosidase	pale, faintly striated inclusions		X	Canine ^{39,40} Silky terrier
Inherited Copper Toxicosis	copper	COMMD-1 ATPB-7/ATPA-7	granules (blue-grey, yellow-brown, red-brown)	X	X (secondary)	Canine (see Chapter 7 copper associated chronic hepatitis)
Mannosidosis	glycoprotein derived mannose-rich oligosaccharides	alpha or beta mannosidase	clear vacuoles	X	X	Canine German shepherd dog ⁶⁸ Feline ⁴¹⁻⁴³ Persian, DSH, DLH
Mucolipidosis type II	mucopolysaccharids, lipids, glycoproteins	N-acetyl glucosamine-1-phosphotransferase	clear vacuoles		X (Fibroblasts /endothelial lining cells)	Feline ⁴⁴ DSH
Mucopolysaccharidosis type I	heparan and dermatan sulphate	alpha-L-iduronidase	clear vacuoles	X	X	Canine ^{45,46} Plott hound Feline ^{45,47} DSH
Mucopolysaccharidosis type II	heparan and dermatan sulphate	alpha-L-iduronidase sulphatase	clear vacuoles	X	X	Canine ⁴⁸ Labrador retriever
Mucopolysaccharidosis type III	heparan sulphate	heparan sulphatase	clear vacuoles	X	X	Canine ⁴⁹⁻⁵¹ Wirehaired dachshund, New Zealand huntaway dog

Mucopolysaccharidosis type VI	dermatan sulphate	arylsulphatase B	clear vacuoles	X	X	Canine ^{16,52,64} Miniature schnauzer, Miniature pinscher, Miniature poodle Feline ^{16,45,53} DSH, Siamese
Mucopolysaccharidosis type VII	chondroitin and dermatan sulphate	beta-glucuronidase	clear vacuoles	X	X	Canine ^{45,54} Mixed breed Feline ^{55,56,66} DSH
Primary hyperlipoproteinaemia	lipids	lipoprotein lipase	clear vacuoles	X	X (including xanthomata)	Feline ^{57,58}
Sphingomyelin-cholesterol lipidosis (Niemann-Pick disease type C)	unesterified cholesterol	acidic sphingomyelinase	clear vacuoles		X	Canine ^{59,60} Miniature poodle, Boxer Feline ^{61,62,67} DSH, Siamese,

These changes are usually non-specific and can be better evaluated in frozen or plastic-embedded sections using conventional, lectin- and immunohistochemistry or electron microscopy. However, diagnosis depends on either the identification of the storage product or the enzyme deficiency by biochemical methods, or by identification of the genetic defect. Although the changes may be insufficient to cause hepatocellular or Kupffer cell necrosis, particularly severe hepatocellular storage may result in hepatocellular necrosis and inflammation and even chronic hepatitis and cirrhosis may develop.

Erythropoietic protoporphyria

Erythropoietic protoporphyria is known in dogs as an acquired storage disease following administration of a number of xenobiotics among others griseofulvin and the antiarthritic drug 3-[2-(2,4,6-trimethylphenyl)-thiothyl]methylsydnoneis (TTMS).⁽⁸⁾ Drug induced protoporphyria is a result of inhibition of ferrochelatase, compensatory stimulation of the first enzyme in the 5-aminolevulate synthase pathway and massive accumulation of the substrate protoporphyrin. Protoporphyrin is recognized as dark brown pigment in canaliculi and interlobular bile ducts, and in more severe cases also in Kupffer cells and macrophages (Fig. 13). The pigment displays bright red birefringence with a centrally located dark Maltese cross. Occasionally severe deposition of the pigment may result in chronic hepatitis and cirrhosis^(9, 10).

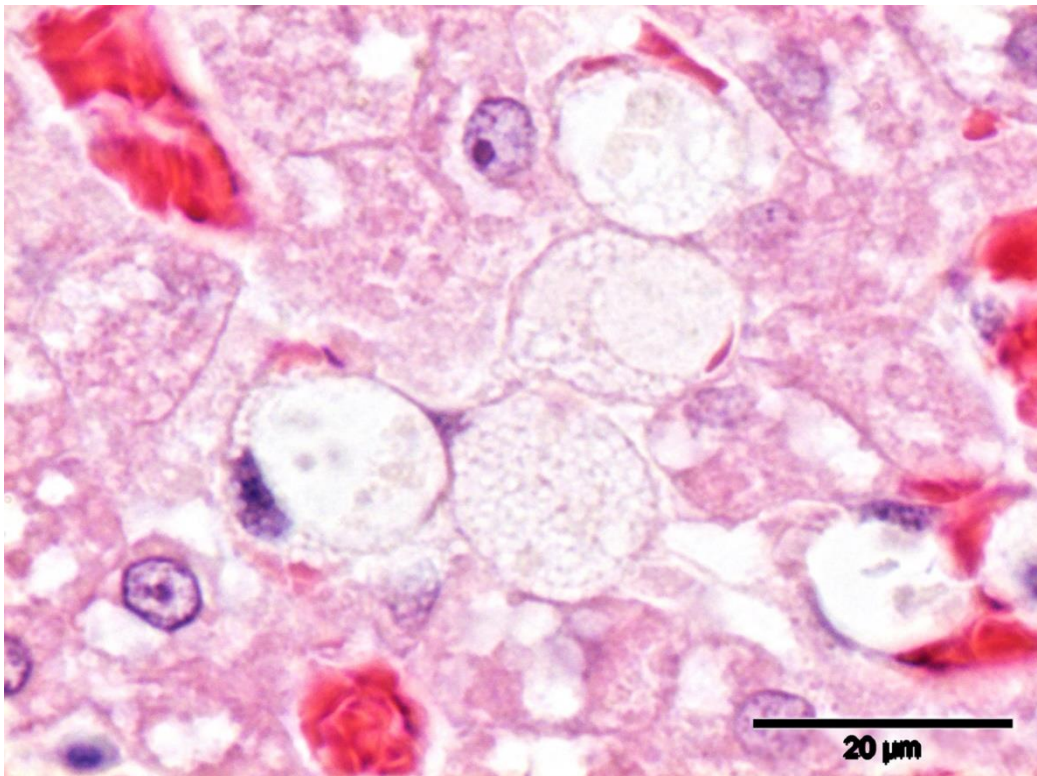


Fig. 11. Dog. Fucosidosis. Swollen Kupffer cells – macrophages with vacuolisation of the cytoplasm. HE.

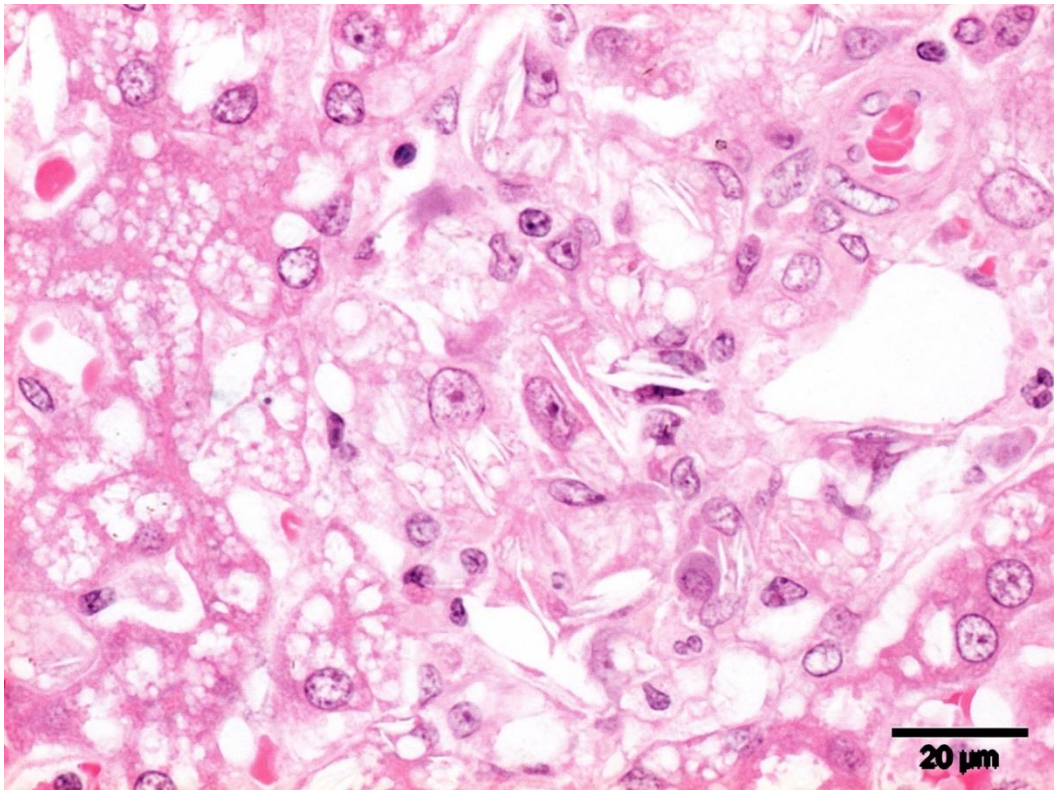


Fig. 12. Dog. Cholesterol ester storage disease (Wolman's disease). Vacuolation of hepatocytes and vacuolation and cholesterol crystals in macrophages in the portal area. HE.

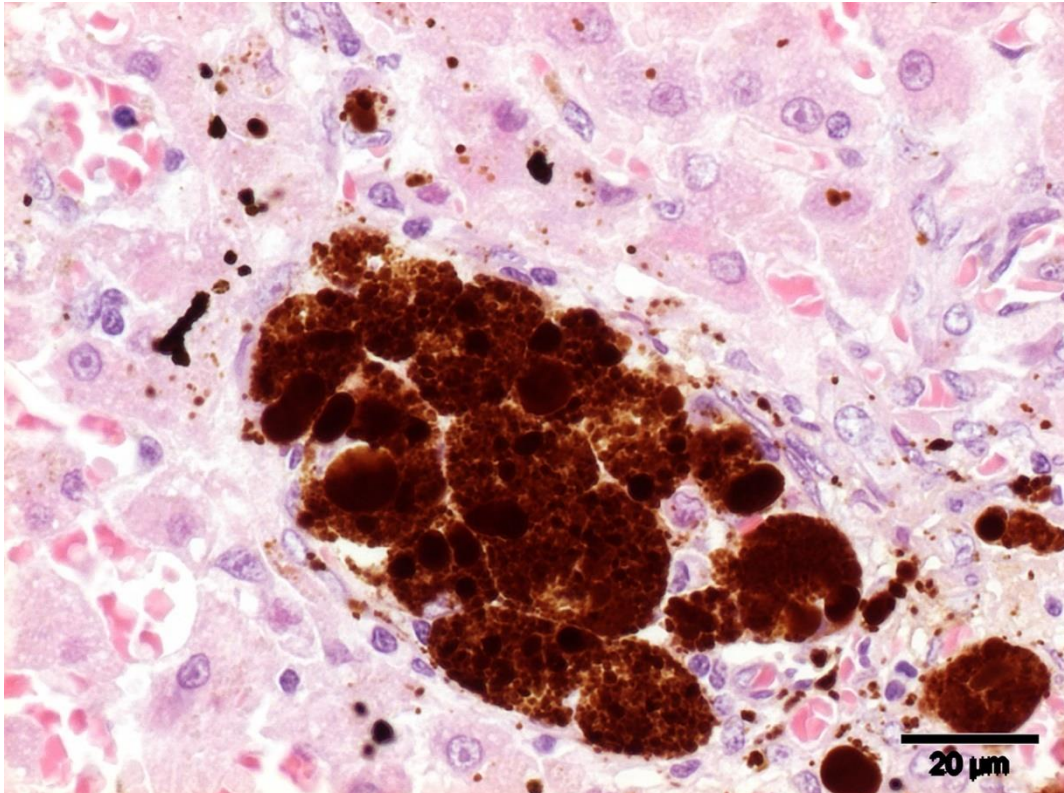


Fig. 13. Dog. Erythropoietic protoporphyria. Darkbrown pigment (protoporphyrin) in canaliculi and in macrophages. HE.

MISCELLANEOUS CONDITIONS

Hepatocytes

Cytoplasmic Alterations

- **Protein droplets** are variably sized eosinophilic (hyaline) droplets in the cytoplasm of hepatocytes (Fig. 14). They may consist of engulfed serum proteins and represent a non-specific and variable finding associated with shock, ischemia, or other acute hepatocellular injury. They also may consist of fibrinogen and acute phase proteins (alpha-1-antitrypsin, haptoglobin, alpha-macroglobulin) retained in the endoplasmic reticulum in hepatocytes as seen in dogs with chronic hepatitis and cirrhosis⁽¹¹⁾
- **Eosinophilic cytoplasmic bodies** of variable size may be the result of phagocytosis of apoptotic bodies by hepatocytes
- **'Ground glass' appearance:** swollen hepatocytes with pink finely granular to hyaline cytoplasm (Fig. 15) associated with an increase in the smooth endoplasmic reticulum and induction of the hepatic microsomal drug metabolizing enzymes. Ground glass appearance of hepatocytes may be caused by drugs (e.g. phenobarbital) and other substances that cause induction of cytochrome P450 enzymes
- **Lipofuscin:** brown-yellow pigmented granular material in hepatocytes within lysosomes and most abundant in a pericanalicular location (Fig. 16). It is present in normal livers in varying amount. Increased hepatocellular lipofuscin occurs with age, particularly in cats, and is most prominent in the centrilobular hepatocytes.
- **Accumulation of copper** is seen as gray-yellow to gray-brown or gray-blue granules in the cytoplasm. The color of the granules may vary depending on tissue preparation procedures used by the laboratory (Fig. 17). Copper can be specifically identified by histochemical stains (e.g. rubeanic acid or rhodanine for copper and orcein for copper-binding protein). Storage of copper can be primary due to a genetic defect in one of the genes associated with copper metabolism, such as in inherited copper toxicosis in Bedlington terriers or secondary due to excessive copper intake. Copper accumulation in hepatocytes is also a hallmark of cholate-stasis (Chapter 5); however, with persistent cholestasis such as in chronic extrahepatic cholestasis and chronic hepatitis/cirrhosis the liver has normal copper concentrations and genes related to copper metabolism remain at concentration similar to normal dogs⁽⁷²⁾.
- **Accumulation of iron** is seen as brown granules in the cytoplasm of hepatocytes (Fig. 18, 19) and may be seen with increased red blood cell turnover, anaemia of chronic disease and after administration of iron.
- **Emperipolesis** is a rare phenomenon of invagination of the cell membrane and engulfment of complete cells. It is particularly seen in epitheliotropic T-cell lymphoma (Fig. 20), where the neoplastic lymphocytes are engulfed by the hepatocytes.⁽¹²⁾

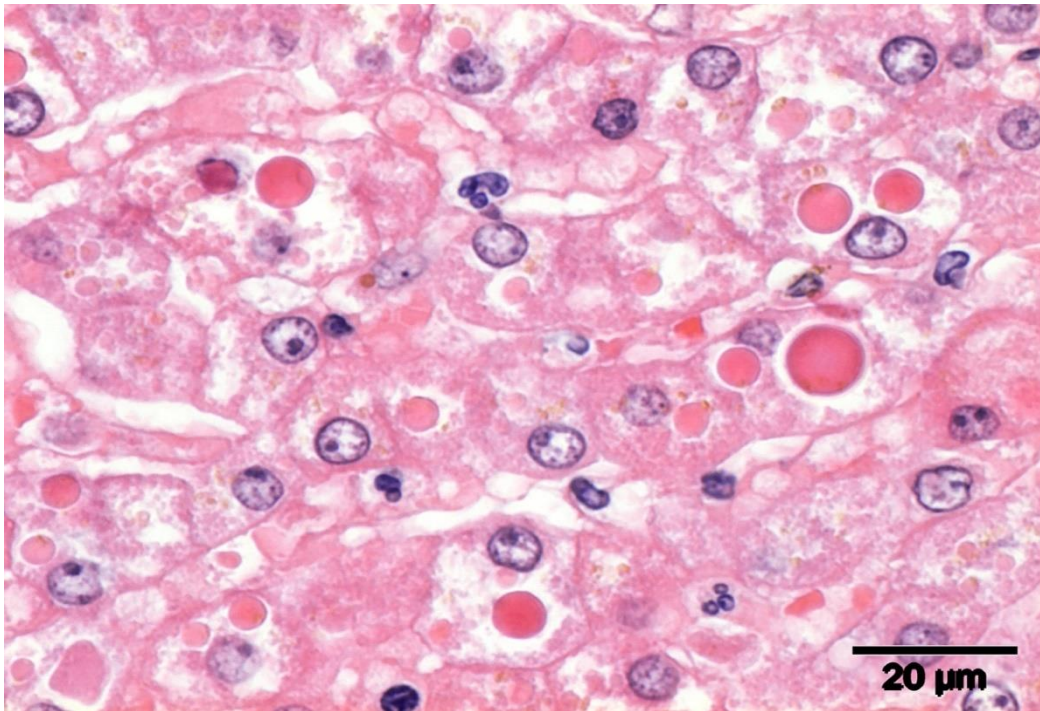


Fig. 14. Dog. Protein droplets (engulfed serum proteins) in the hepatocytic cytoplasm. HE.

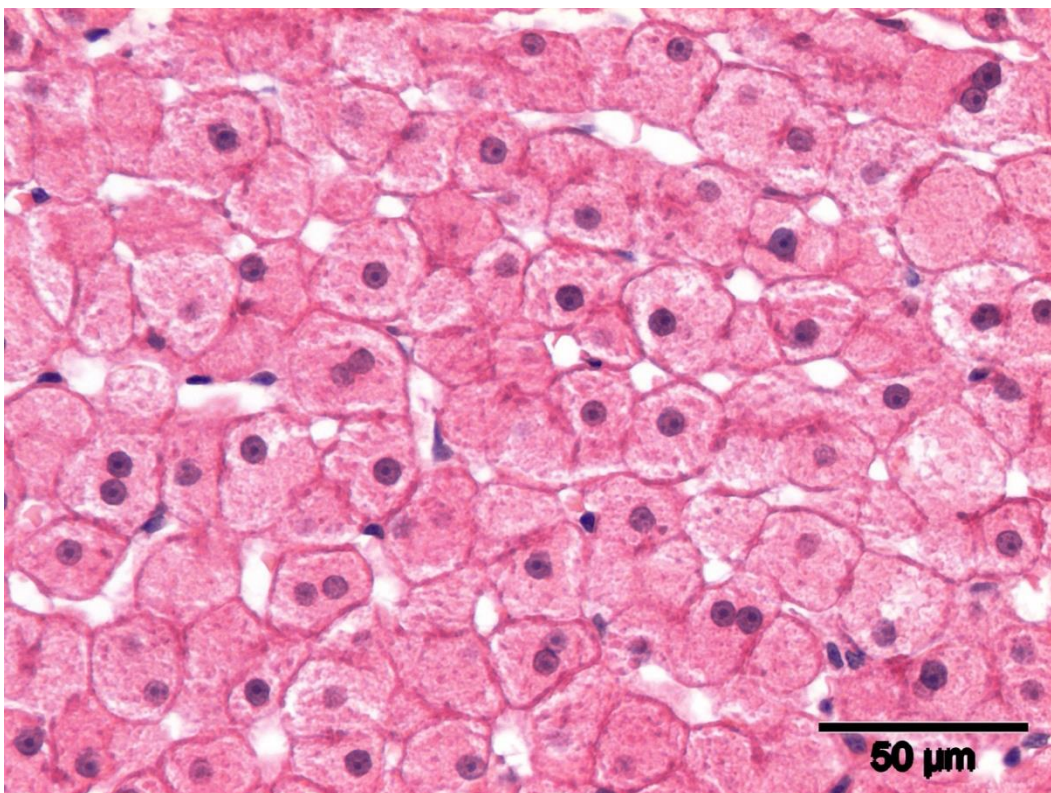


Fig. 15. Dog. Ground glass appearance associated with phenobarbital medication. HE.

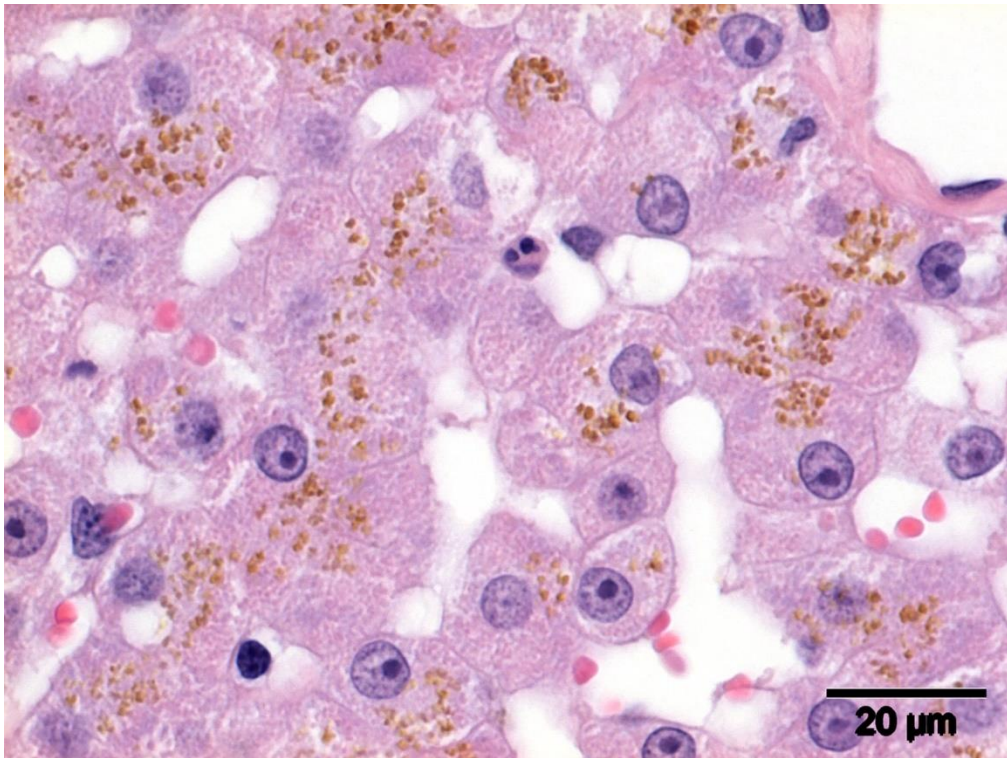


Fig. 16. Cat. Lipofuscin deposition in a pericanalicular location in centrilobular hepatocytes. HE.

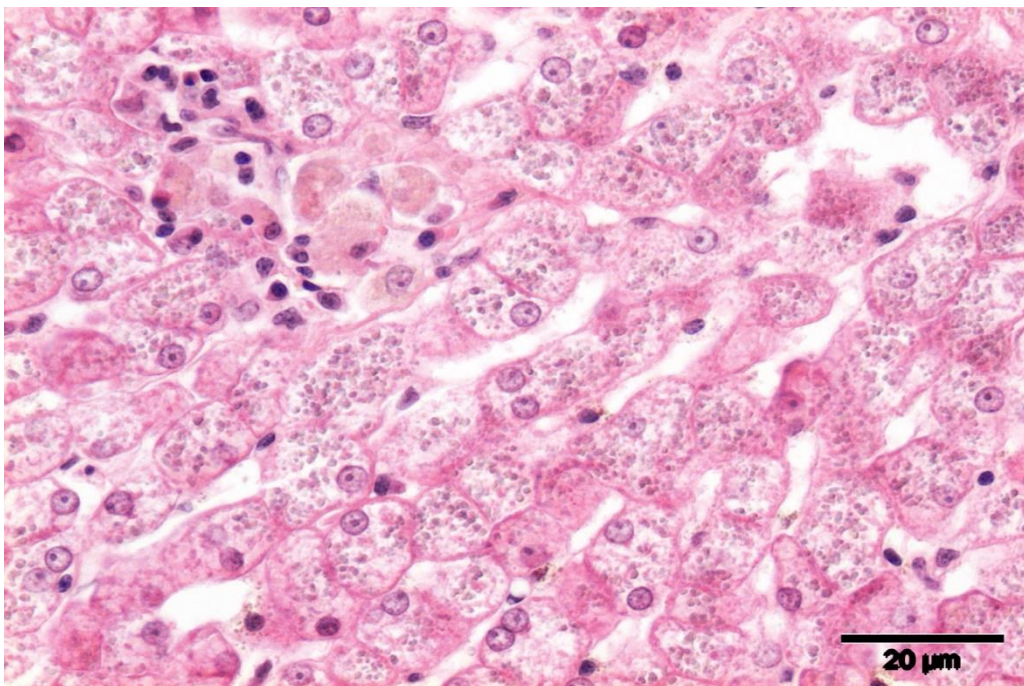


Fig. 17. Bedlington terrier. Gray-yellow copper containing granules diffusely dispersed in the hepatocytic cytoplasm. Inherited copper toxicosis. HE.

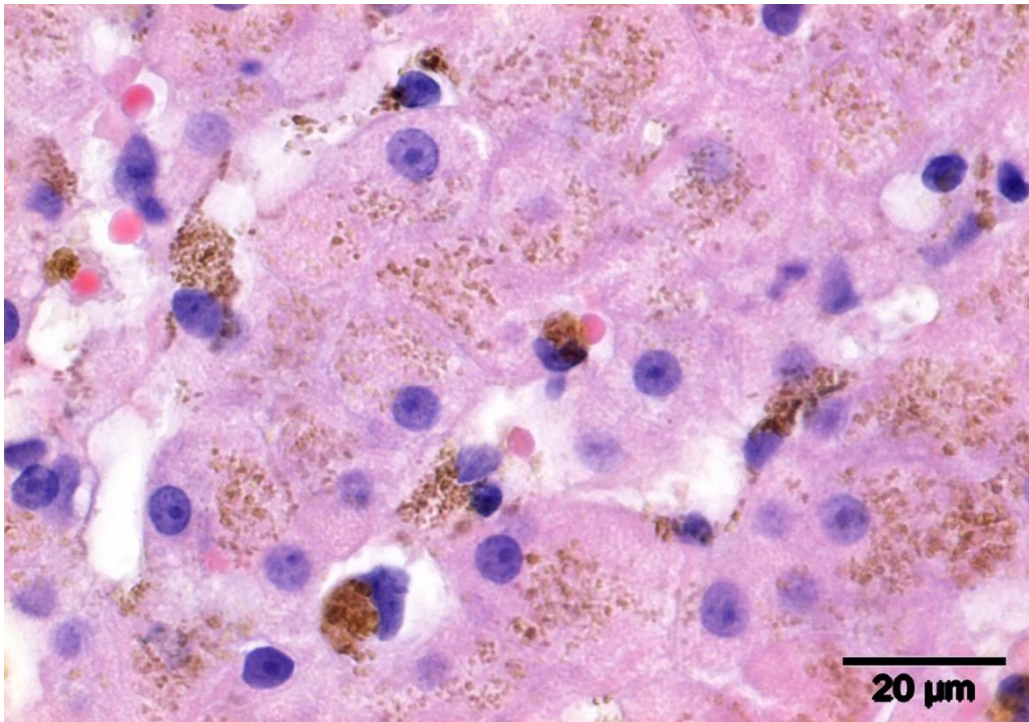


Fig. 18. Cat. Brown iron containing granules in the cytoplasm of hepatocytes and Kupffer cells. Erythremic myelosis. HE.

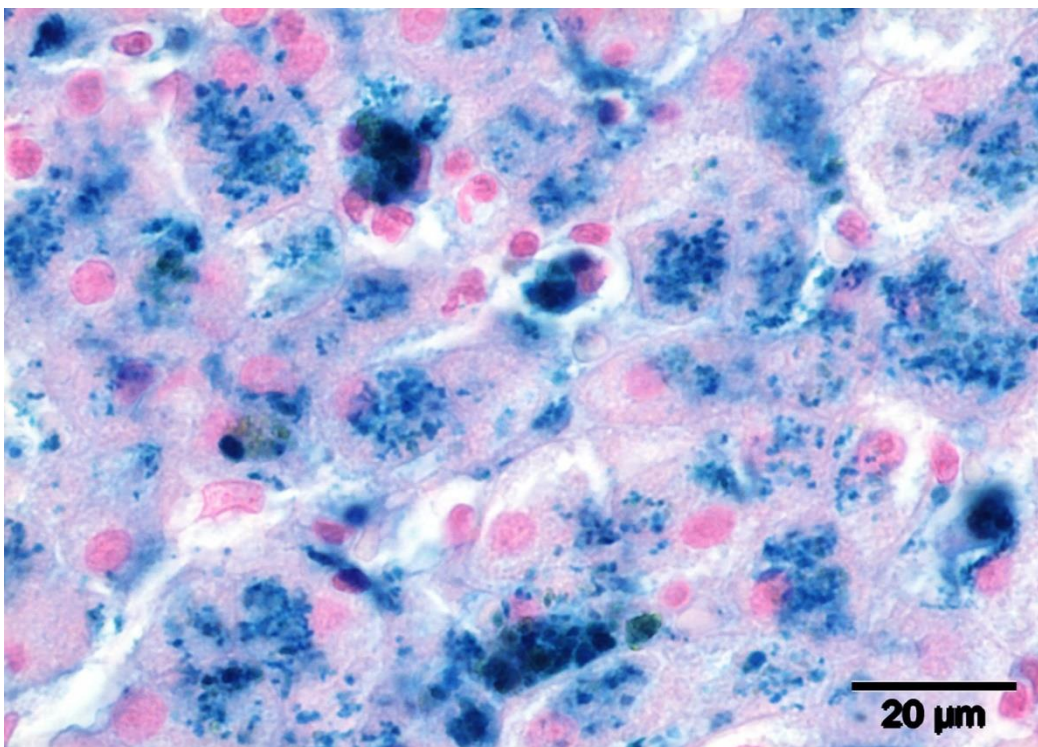


Fig. 19. Cat (same animal as Fig 8.18). Blue staining iron pigment in hepatocytes and Kupffer cells. Prussian blue stain for iron.

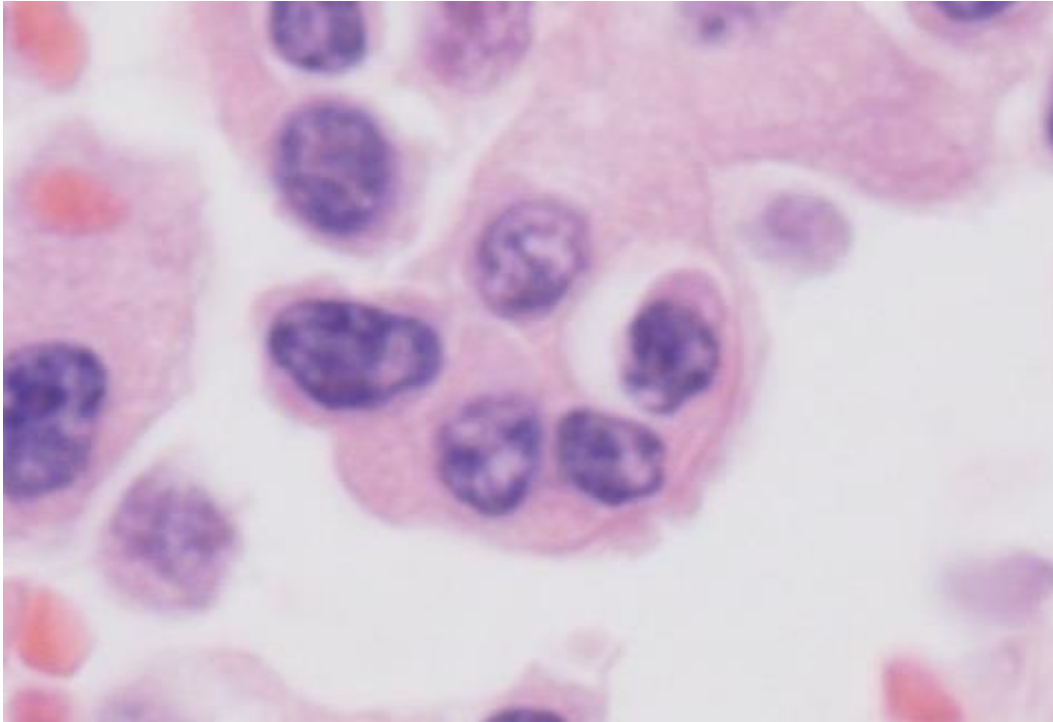


Fig. 20. Emperipolesis. Engulfment of neoplastic T lymphocytes by hepatocytes. HE

Nuclear Alterations

- **'Brick' inclusions** (Fig. 21) are rectangular to rhomboid brightly eosinophilic bodies in the nuclei of hepatocytes in dogs that are of no known cause of significance.
- **Cytoplasmic invaginations** occur as round to oval protrusions of the cytoplasm into the nucleus and are of no known significance
- **Glycogen inclusions** (Fig. 22) occur as clear inclusions in the nucleus and are of unknown significance. They are non-specific and may occur among others in diabetes mellitus and hepatocellular neoplasia
- **Intranuclear viral inclusions** (Fig. 23) may be observed in various viral infections (see hepatitis)
- **Intranuclear acid fast inclusions** have been reported in dogs with lead intoxication but are extremely rare.⁽¹³⁾

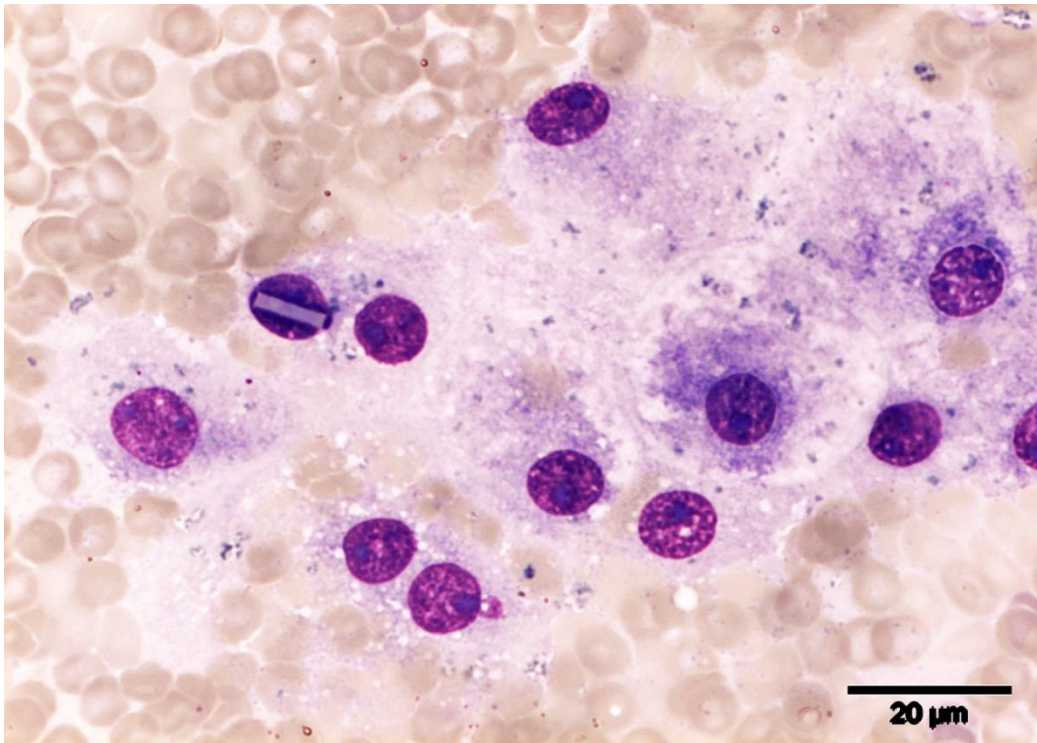


Fig. 21. Dog. Cytological specimen. Intranuclear brick inclusion. May-Grünwald Giemsa stain.

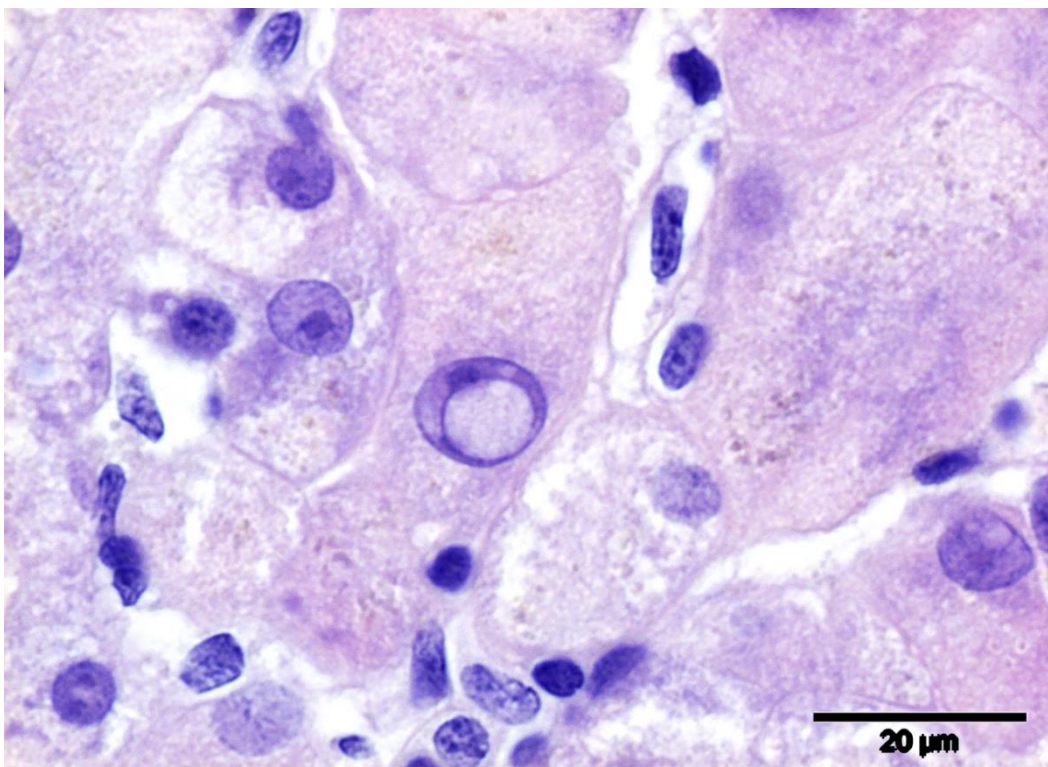


Fig. 22. Dog. Intranuclear glycogen inclusion. HE.

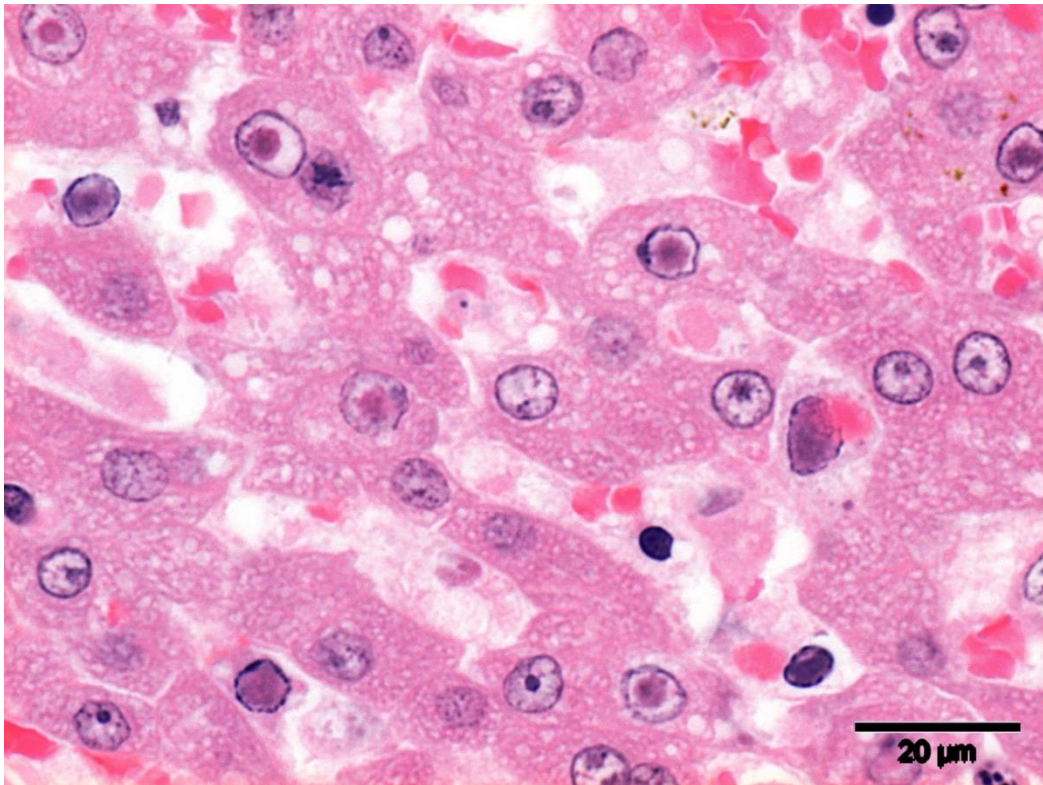


Fig. 23. Dog. Basophilic and amphophilic intranuclear viral inclusions in hepatocytes and endothelial cells. Adenovirus type 1 infection.

Hepatic stellate cells

Hepatic stellate cells have large lipid vacuoles. Increased numbers of hepatic stellate cells are observed in older cats and less pronounced in old dogs. They are recognized as solitary cells with a single large empty vacuole with displacement of the nucleus to the periphery of the cell (Fig. 24). The significance of this change is not known.

Chronic vitamin A intoxication is known in cats and causes hypertrophy and hyperplasia of lipid-laden hepatic stellate cells (Fig. 25), which show specific vitamin A autofluorescence, as well as sinusoidal fibrosis^(69,70,71) with or without mild hepatocellular steatosis.

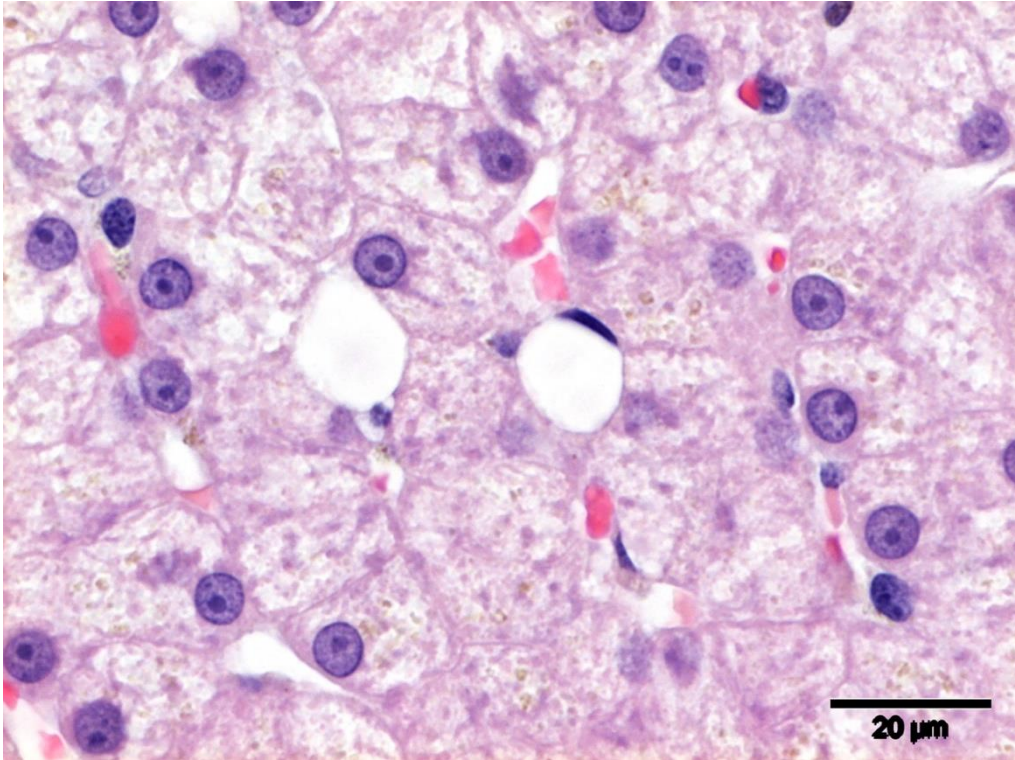


Fig. 24. Dog. Hepatic stellate cell with a large lipid vacuole and displacement of the nucleus to the periphery of the cell. HE.

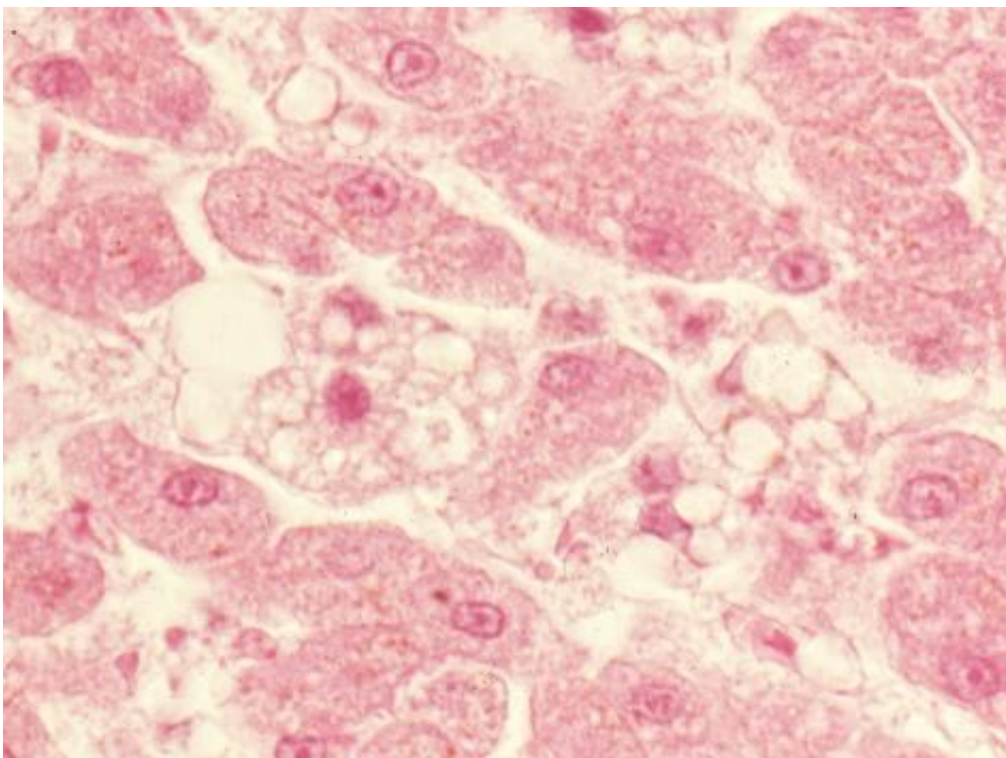


Fig. 25. Cat. Hypertrophy and hyperplasia of lipid laden hepatic stellate cells. Vitamin A intoxication. HE. (From Mouwen JMVM, De Groot ECBM. Atlas of Veterinary Pathology. Utrecht: Bunge; 1982, with permission).

Kupffer cells

- **Necrosis** of Kupffer cells may be seen in sepsis and toxemia.
- **Erythrophagocytosis / hemosiderosis** (Fig. 26) is observed in conditions with increased red cell turnover, such as hemolytic anemia, anaemia of chronic disease and chronic hepatitis. The Kupffer cells may contain erythrocytes (erythrophagocytosis) and, or increased amounts of iron pigment (hemosiderosis)
- **Bile pigment** (Fig. 27) can be seen in cases with marked cholestasis and will be present as yellow-brown to yellow-green intracytoplasmic granules or bile plugs.
- **Eosinophilic cytoplasmic bodies** of variable size may be the result of phagocytosis of fragments of apoptotic cells by Kupffer cells.
- **Ceroid** is a yellow-brown, PAS positive, lipid breakdown product that accumulates in Kupffer cells and macrophages as a result of increased hepatocyte turnover due to necrosis / apoptosis. (Fig. 28)

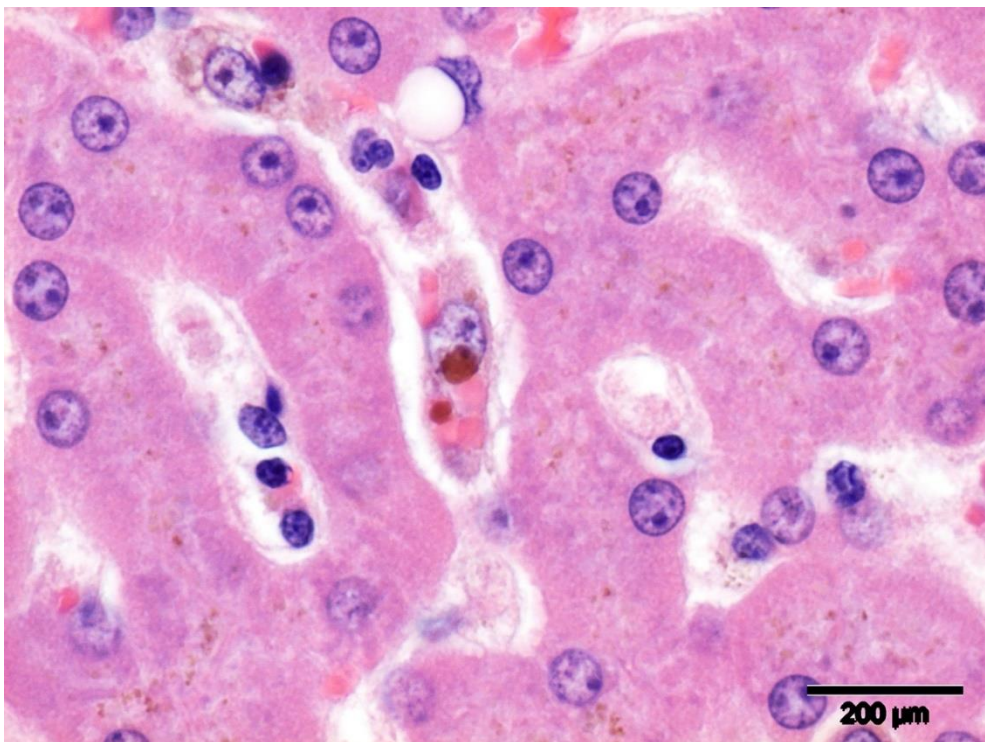


Fig. 26. Dog. Erythrophagocytosis and haemosiderosis of Kupffer cell. Immune-mediated haemolytic anaemia. HE.

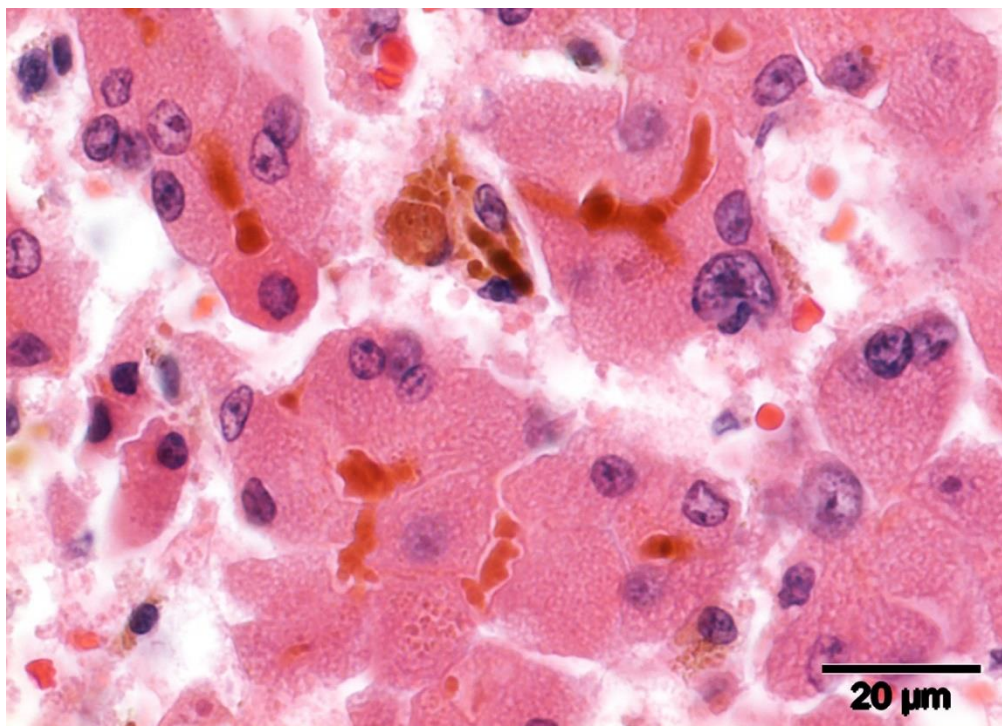


Fig. 27. Dog. Bile plugs in canaliculi and phagocytosed bile plugs in Kupffer cells. HE.

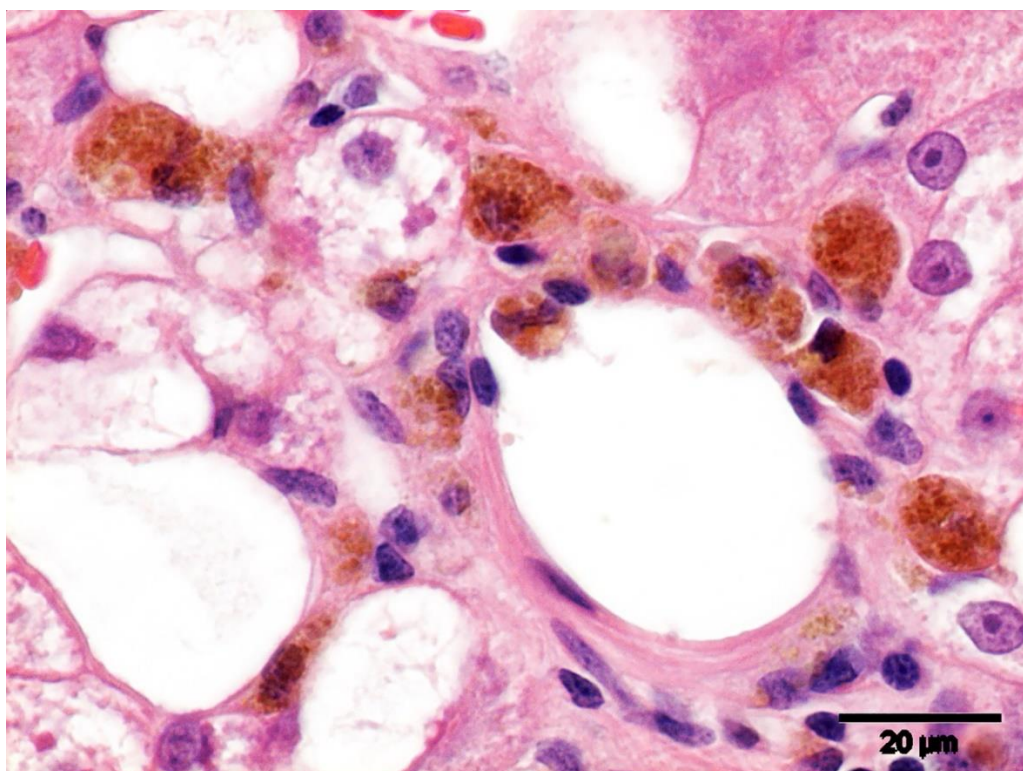


Fig. 28. Dog. Ceroid containing macrophages. HE.

Lipogranulomas and pigment granulomas

Lipogranulomas (syn : fatty cysts⁽¹⁴⁾) and pigment granulomas are poorly defined lesions of undetermined significance with overlapping features. Lipogranulomas are aggregates of

ceroid-laden, foamy fat-containing macrophages and sometimes also some lymphocytes and plasma cells (Fig. 29). They probably are associated with previous hepatocyte death and occur both in the parenchyma as well as in the portal and perivenular connective tissue. Pigment granulomas generally consist of both ceroid- and iron pigment containing macrophages (Fig. 30); often also lymphocytes or plasma cells are present. They are present throughout the parenchyma and are regularly seen particularly in older animals without known significance.

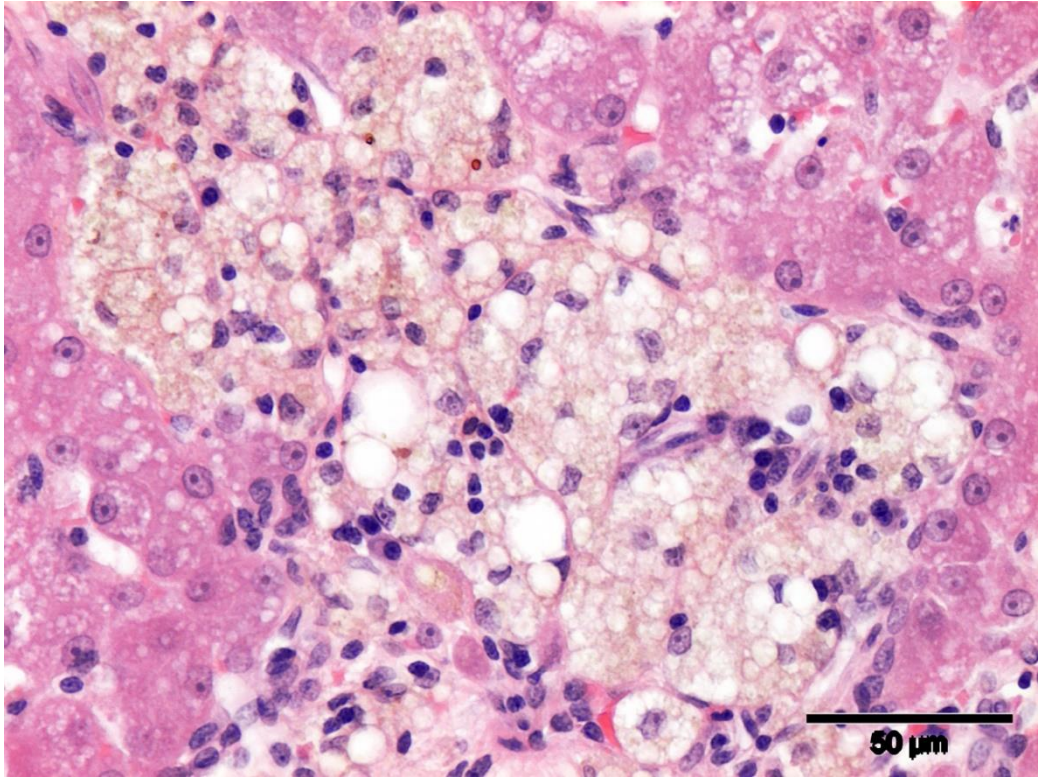


Fig. 29 Dog. Lipogranuloma. HE.

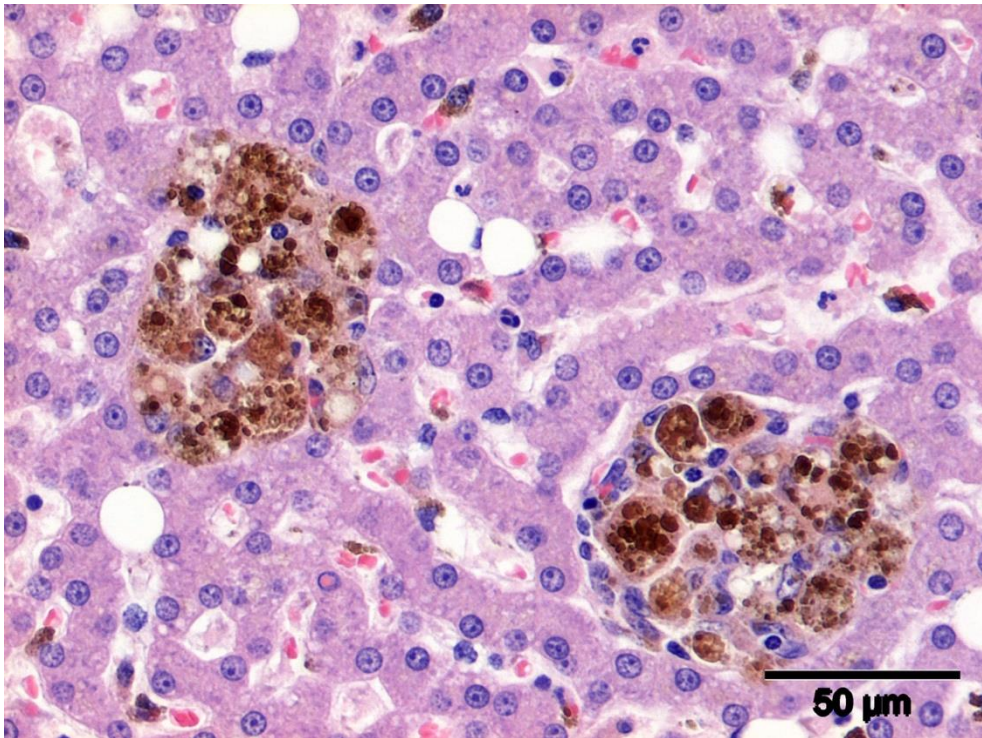


Fig. 30. Dog. Pigment granulomas. HE.

Extramedullary hemopoiesis

In both canine and feline neonates **extramedullary hemopoiesis** of all hematopoietic elements is a normal condition. Whereas the erythropoietic activity (Fig. 31) and megakaryocytes are particularly seen in the parenchyma, the myelopoiesis is mainly seen in the stromal tissue of the portal and perivenular areas. In septicemic bacterial diseases the myelopoietic activity may be markedly increased.

In older dogs and less frequent in cats **extramedullary erythropoiesis and megakaryocytes** can be observed in immune-mediated haemolytic anaemia and thrombocytopenia.

Extramedullary myelopoiesis (Fig. 32) is regularly seen as small foci of band and segmented neutrophils in dogs with steroid induced hepatopathy, and in chronic purulent inflammation as pyometra and pleuritis or peritonitis due to nocardiosis or actinomycosis.

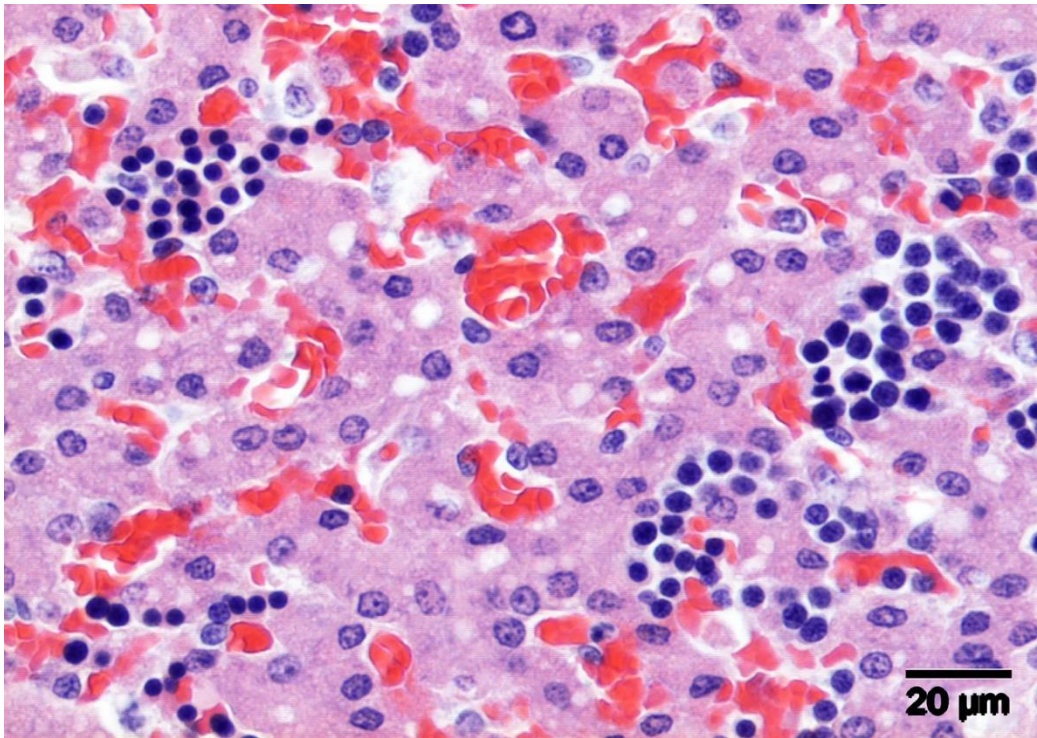


Fig. 31. Dog, neonate. Extramedullary erythropoiesis. HE.

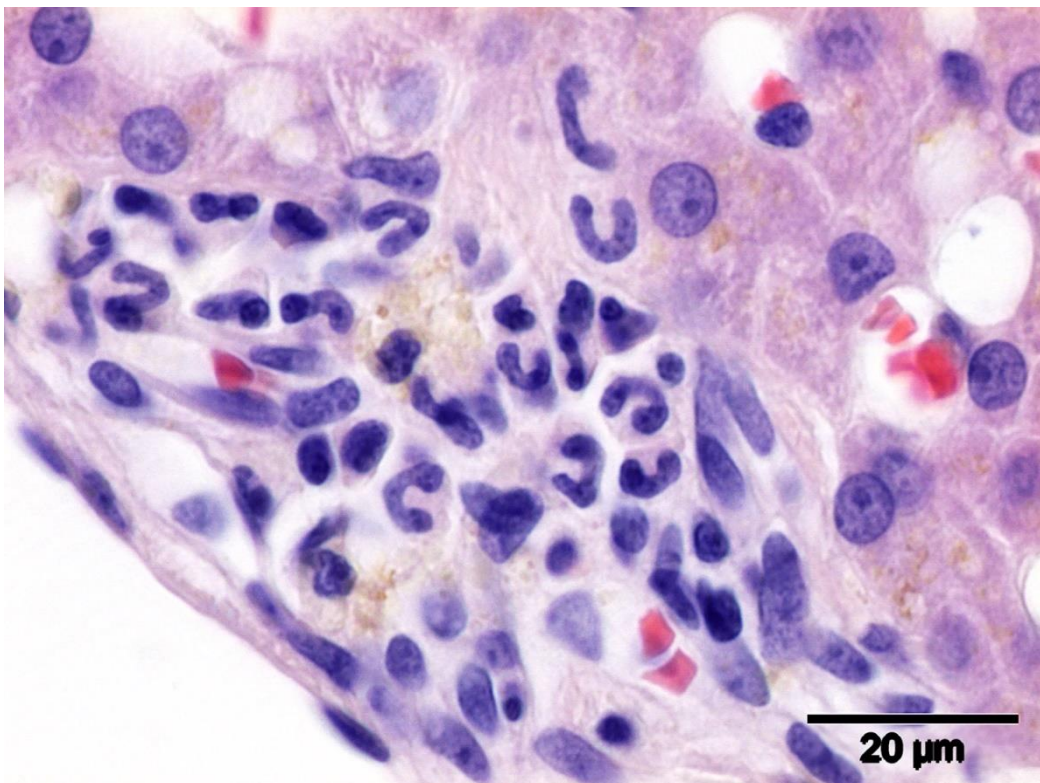


Fig. 32. Dog. Extramedullary myelopoiesis. HE.

REFERENCES

1. Farrar ET, Washabau RJ, Saunders HM. Hepatic abscesses in dogs: 14 cases (1982-1994). *J Am Vet Med Assoc* 1996;208(2):243-247.
2. Sergeeff JS, Armstrong PJ, Bunch SE. Hepatic abscesses in cats: 14 cases (1985 - 2002). *J Vet Intern Med* 2004;18:295-300.
3. Cotran RS, Kumar V, Collins T. Robbins: Pathologic Basis of Disease. Philadelphia: W.B.Saunders Company; 2000.
4. Rubarth S. Leber und Gallenwege. In: Dobberstein J, Pallaske G, Stünzi H, editors. *Joest-Handbuch der Speziellen Pathologischen Anatomie der Haustiere*. 3rd ed. Berlin: Paul Parey Verlag; 1967. p. 1-177.
5. Slappendel RJ, Ferrer L. Leishmaniasis. In: Greene CE, editor. *Infectious diseases of the dog and cat*. 2nd ed. Philadelphia: W.B. Saunders Company; 1998. p. 450-458.
6. Kier AB, Greene CE. Cytauxzoonosis. In: Greene CE, editor. *Infectious diseases of the dog and cat*. 2nd ed. Philadelphia: W.B. Saunders Company; 1998. p. 470-473.
7. Gillespie TN, Washabau RJ, Goldschmidt MH, et al. Detection of *Bartonella henselae* and *Bartonella clarridgeiae* DNA in hepatic specimens from two dogs with hepatic disease. *J Am Vet Med Assoc* 2003;222:47-51.
8. Greaves P. *Histopathology of preclinical toxicity studies*. 2nd ed. Amsterdam: Elsevier; 2000.
9. Greydanus-van der Putten SW, van Esch E, Kamerman J et al. Drug-induced protoporphyria in Beagle dogs. *Toxicol Pathol* 33:2005:720–725.
10. Kroeze EJ, Zentek J, Edixhoven - Bosdijk A et al. Transient erythropoietic protoporphyria associated with chronic hepatitis and cirrhosis in a cohort of German shepherd dogs. *Vet Rec* 2006;. 158:120-124.
11. Vatne M, Andersson, M., Sevelius, E et al. Immunohistochemical investigation of four glycoproteins in the hepatocytes of dogs with chronic liver disease. *European J. Vet. Pathol.* 2001;7:51-59.
12. Ossent P, Stockli RM, Pospischil A. Emperipolesis of lymphoid neoplastic cells in feline hepatocytes. *Vet Pathol* 1989;26(3):279-80.
13. Jones TC, Hunt RD, King NW. *Veterinary Pathology*. 6th ed. Baltimore: Williams and Wilkins; 1997.
14. Kelly WR. The liver and biliary system. In: Jubb KVF, Kennedy PC, Palmer N, editors. *Pathology of domestic animals*. 4th ed. San Diego: Academic Press; 1992. p. 319-406.
15. Taylor RM, Farrow BR. Ceroid-lipofuscinosis in border collie dogs. *Acta Neuropathol (Berl)* 1988;75:627-31.

16. Warren CD, Alroy J. Morphological, biochemical and molecular biology approaches for the diagnosis of lysosomal storage diseases. *J Vet Diagn Invest* 2000;12:483-96.
17. Jolly RD, Walkley SU. Lysosomal storage diseases of animals: an essay in comparative pathology. *Vet Pathol* 1997;34:527-548.
18. Nakayama H, Uchida K, Shouda T, et al. Systemic ceroid-lipofuscinosis in a Japanese domestic cat. *J Vet Med Sci* 1993;55:829-831.
19. Hänichen T, Breuer, W. and Hermanns, W. Canine lipid storage disease. *Eur J Vet Path* 1995;1:37-44.
20. Keller CB, Lamarre J. Inherited lysosomal storage disease in an English springer spaniel. *J Am Vet Med Assoc* 1992;200:194-195.
21. Herrtage ME, Palmer AC, Blakemore WF. Canine fucosidosis. *Vet Rec* 1985;117:451-452.
22. Knowles K, Alroy J, Castagnaro M, et al. Adult-onset lysosomal storage disease in a Schipperke dog: clinical, morphological and biochemical studies. *Acta Neuropathol (Berl)* 1993;86:306-312.
23. Muller G, Alldinger S, Moritz A, et al GM1-gangliosidosis in Alaskan huskies: clinical and pathologic findings. *Vet Pathol* 2001;38:281-290.
24. Alroy J, Orgad U, Ucci AA, et al. Neurovisceral and skeletal GM1-gangliosidosis in dogs with beta-galactosidase deficiency. *Science* 1985;229:470-472.
25. Rodriguez M, O'Brien JS, Garrett RS, Powell HC. Canine GM1 gangliosidosis. An ultrastructural and biochemical study. *J Neuropathol Exp Neurol* 1982;41:618-629.
26. Saunders GK, Wood PA, Myers RK, et al. GM1 gangliosidosis in Portuguese water dogs: pathologic and biochemical findings. *Vet Pathol* 1988;25:265-269.
27. Shell LG, Potthoff AI, Carithers R, et al Katherman A, Saunders GK, Wood PA, et al. Neuronal-visceral GM1 gangliosidosis in Portuguese water dogs. *J Vet Intern Med* 1989;3:1-7.
28. Yamato O, Ochiai K, Masuoka Y, Hayashida E, Tajima M, Omae S, et al. GM1 gangliosidosis in shiba dogs. *Vet Rec* 2000;146:493-496.
29. De Maria R, Divari S, Bo S, et al. Beta-galactosidase deficiency in a Korat cat: a new form of feline GM1-gangliosidosis. *Acta Neuropathol (Berl)* 1998;96:307-314.
30. Baker HJ, Walkley SU, Rattazzi MC, et al. Feline gangliosidoses as models of human lysosomal storage diseases. *Prog Clin Biol Res* 1982;94:203-212.
31. Cummings JF, Wood PA, Walkley SU, et al. GM2 gangliosidosis in a Japanese spaniel. *Acta Neuropathol (Berl)* 1985;67:247-53.

32. Yamato O, Matsuki N, Satoh H, et al. Sandhoff disease in a golden retriever dog. *J Inher Metab Dis* 2002;25:319-320.
33. Karbe EaS, B. Familial amaurotic idiocy in male German Shorthair Pointers. *Vet Pathol* 1967;6:223-232.
34. Cork LC, Munnell JF, Lorenz MD. The pathology of feline GM2 gangliosidosis. *Am J Pathol* 1978;90:723-734.
35. Brix AE, Howerth EW, McConkie-Rosell A, et al. Glycogen storage disease type Ia in two littermate Maltese puppies. *Vet Pathol* 1995;32:460-465.
36. Ceh L, Hauge JG, Svenkerud R, Strande A. Glycogenosis type III in the dog. *Acta Vet Scand* 1976;17:210-222.
37. Rafiquzzaman M, Svenkerud, R., Strande, A. and Hauge, J.G. Glycogenosis in the dog. *Acta Vet Scand* 1976;17:196-209.
38. Fyfe JC, Giger, U., Van Winkle, TJ, et al. D.F. Glycogen storage disease type iv: inherited deficiency of branching enzyme activity in cats. *Pediatr. Res* 1992;32:719-725.
39. Hartley WJ, Blakemore WF. Neurovisceral glucocerebroside storage (Gaucher's disease) in a dog. *Vet Pathol* 1973;10:191-201.
40. Farrow BR, Hartley WJ, Pollard AC, et al. Gaucher disease in the dog. *Prog Clin Biol Res* 1982;95:645-653.
41. Cummings JF, Wood PA, de Lahunta A, et al. The clinical and pathologic heterogeneity of feline alpha-mannosidosis. *J Vet Intern Med* 1988;2(4):163-170.
42. Jezyk PF, Haskins ME, Newman LR. Alpha-mannosidosis in a Persian cat. *J Am Vet Med Assoc* 1986;189:1483-1485.
43. Maenhout T, Kint JA, Dacremont G, et al. Mannosidosis in a litter of Persian cats. *Vet Rec* 1988;122:351-354.
44. Hubler M, Haskins ME, Arnold S, et al. Mucopolidosis type II in a domestic shorthair cat. *J Small Anim Pract* 1996;37:435-441.
45. Haskins ME, Otis EJ, Hayden JE, et al. Hepatic storage of glycosaminoglycans in feline and canine models of mucopolysaccharidoses I, VI, and VII. *Vet Pathol* 1992;29:112-119.
46. Spellacy E, Shull RM, Constantopoulos G, Neufeld EF. A canine model of human alpha-L-iduronidase deficiency. *Proc Natl Acad Sci U S A* 1983;80:6091-6095.
47. Haskins ME, Aguirre GD, Jezyk PF, et al. The pathology of the feline model of mucopolysaccharidosis I. *Am J Pathol* 1983;112:27-36.

48. Wilkerson MJ, Lewis DC, Marks SL, Prieur DJ. Clinical and morphologic features of mucopolysaccharidosis type II in a dog: naturally occurring model of Hunter syndrome. *Vet Pathol* 1998;35:230-233.
49. Fischer A, Carmichael KP, Munnell JF, et al. Sulfamidase deficiency in a family of Dachshunds: a canine model of mucopolysaccharidosis IIIA (Sanfilippo A). *Pediatr Res* 1998;44:74-82.
50. Jolly RD, Ehrlich PC, Franklin RJ, et al. Histological diagnosis of mucopolysaccharidosis IIIA in a wire-haired dachshund. *Vet Rec* 2001;148:564-567.
51. Yogalingam G, Pollard T, Gliddon B, et al. Identification of a mutation causing mucopolysaccharidosis type IIIA in New Zealand Huntaway dogs. *Genomics* 2002;79:150-153.
52. Neer TM, Dial SM, Pechman R, et al. Clinical vignette. Mucopolysaccharidosis VI in a miniature pinscher. *J Vet Intern Med* 1995;9:429-433.
53. Haskins ME, Aguirre GD, Jezyk PF, Patterson DF. The pathology of the feline model of mucopolysaccharidosis VI. *Am J Pathol* 1980;101:657-674.
54. Haskins ME, Aguirre GD, Jezyk PF, et al. Mucopolysaccharidosis type VII (Sly syndrome). Beta-glucuronidase-deficient mucopolysaccharidosis in the dog. *Am J Pathol* 1991;138:1553-1555.
55. Schultheiss PC, Gardner SA, Owens JM, et al. Mucopolysaccharidosis VII in a cat. *Vet Pathol* 2000;37:502-505.
56. Gitzelmann R, Bosshard NU, Superti-Furga A, et al. Feline mucopolysaccharidosis VII due to beta-glucuronidase deficiency. *Vet Pathol* 1994;31:435-443.
57. Thompson JC, Johnstone AC, Jones BR, Hancock WS. The ultrastructural pathology of five lipoprotein lipase-deficient cats. *J Comp Pathol* 1989;101:251-262.
58. Johnstone AC, Jones BR, Thompson JC, Hancock WS. The pathology of an inherited hyperlipoproteinaemia of cats. *J Comp Pathol* 1990;102:125-137.
59. Bundza A, Lowden JA, Charlton KM. Niemann-Pick disease in a poodle dog. *Vet Pathol* 1979;16:530-538.
60. Kuwamura M, Awakura T, Shimada A, et al. Type C Niemann-Pick disease in a boxer dog. *Acta Neuropathol (Berl)* 1993;85:345-348.
61. Lowenthal AC, Cummings JF, Wenger DA, et al. Feline sphingolipidosis resembling Niemann-Pick disease type C. *Acta Neuropathol (Berl)* 1990;81:189-197.
62. Wenger DA, Sattler, M., Kudoh, T, et al. Niemann-Pick disease: a genetic model in Siamese cats. *Science* 1980;208:1470-1473.

63. Gregory BL, Shelton GD, Bali DS et al. Glycogen storage disease type IIIa in curly-coated retrievers. *J Vet Intern Med* 2007;21:40-46.
64. Jolly RD, Hopwood JJ, Marshall NR et al. Mucopolysaccharidosis type VI in a Miniature Poodle-type dog caused by a deletion in the arylsulphatase B gene. *N Z Vet J.* 2012;60:183-188.
65. Brooks ED, Yi H, Stephanie L Austin SL, et al. Natural Progression of Canine Glycogen Storage Disease Type IIIa.. *Comp Med* 2016;66:41–51.
66. Wang P, Sorenson J, Strickland S et al. Mucopolysaccharidosis VII in a Cat Caused by 2 Adjacent Missense Mutations in the GUSB Gene. *J Vet Intern Med* 2015;29:1022–1028.
67. Zampieri S, Bianchi E, Cantile C, et al Characterization of a Spontaneous Novel Mutation in the NPC2 Gene in a Cat Affected by Niemann Pick Type C Disease. *PLoS ONE* 9(11): e112503. doi:10.1371/journal.pone.0112503.
68. Jolly RD, Dittmer KE, Garrick DJ et al. β -Mannosidosis in German Shepherd Dogs. *Vet Pathol* 2019;56:743-748.
69. Guerra JM, Daniel AGT, Thiago P A Aloia TPA et al. Hypervitaminosis A-induced Hepatic Fibrosis in a Cat., *J Feline Med Surg* 2014;16:243-248.
70. Corbee RJ, Tryfonidou MA, Grinwis GCM, et al. Skeletal and hepatic changes induced by chronic vitamin A supplementation in cats. *Vet J* 2014;202:503-509.
71. Senoo H, Imai K, Mezaki Y, et al. Accumulation of Vitamin A in the Hepatic Stellate Cell of Arctic Top Predators. *Anat Rec* 2012;295:1660–1668.
72. Spee B, Arends B, van den Ingh TSGAM et al. Copper Metabolism and Oxidative Stress in Chronic Inflammatory and Cholestatic Liver Diseases in Dogs. *J Vet Intern Med* 2006;20:1085–1092